

AUA Cookie Policy and Agreement

We use cookies and other tracking technologies to improve your browsing experience on our website, to show you personalized content and targeted ads, to analyze our website traffic, and to understand where our visitors are coming from in an effort to continue to serve your needs.

[I agree](#)

[I decline](#)

[Change my preferences](#)

AUA Websites ▾



American
Urological
Association

[Home](#) | [Guidelines & Quality](#) | [Guidelines](#) | [Male Infertility](#)

Guidelines

Diagnosis and Treatment of Infertility in Men: AUA/ASRM Guideline

USING AUA GUIDELINES

This AUA guideline is provided free of charge to the general public for clinical, academic, research and educational use purposes only. Any person or company accessing or using AUA guidelines for promotional or commercial purposes must request a license for use. To request a license for this guideline, please contact Keith Price at kprice@AUAnet.org.

PUBLISHED 2020; AMENDED 2024

[Unabridged version of this Guideline](#) [pdf]

[Guideline Amendment Summary](#) [pdf]

[Algorithm associated with this Guideline](#) [pdf]

To cite this guideline:

Brannigan RE, Hermanson L, Kaczmarek J, Kim SK, Kirkby E, Tanrikut C. Updates to male infertility: AUA/ASRM guideline (2024). J Urol. Published online August 15, 2024. doi:10.1097/JU.0000000000004180.

<https://www.auajournals.org/doi/10.1097/JU.0000000000004180>

Panel Members

Peter N. Schlegel, MD; Mark Sigman, MD; Barbara Collura; Christopher J. De Jonge, PhD; Michael L. Eisenberg, MD; Dolores I. Lamb, PhD; John P. Mulhall, MD; Craig Niederberger MD; Jay I. Sandlow, MD; Rebecca Z. Sokol, MD, MPH; Steven D. Spandorfer, MD; Cigdem Tanrikut, MD; Armand Zini, MD

Amendment Panel

Robert E. Brannigan, MD; Cigdem Tanrikut, MD

Staff and Consultants

Sennett K. Kim; Erin Kirkby, MS; Linnea Hermanson, MA; Janice Kaczmarek, MS; Jeffrey T. Oristaglio, PhD; Jonathan R. Treadwell, PhD

SUMMARY

Purpose

Infertility is due in whole or in part to the male in approximately one-half of all infertile couples. Although many couples can achieve a pregnancy with— intrauterine insemination (IUI) and assisted reproductive technologies (ART) (in vitro fertilization [IVF] with or without intracytoplasmic sperm injection [ICSI]), evaluation of the male is important to most appropriately direct therapy. Some male factor conditions are treatable with medical or surgical therapy, and others may require donor sperm or adoption, if appropriate. Some conditions are life threatening, while others have health and genetic implications for the patient and potential offspring. A male evaluation is necessary to adequately design the management of the patient and the couple. Without an adequate male infertility workup, unnecessary costly, time-consuming, and invasive treatment might be pursued for the female partner.

The purpose of this Guideline is to outline the appropriate evaluation and management of the male partner in an infertile couple. Recommendations proceed from obtaining an appropriate history and physical exam (Appendix I), as well as diagnostic testing, where indicated. Medical therapies, surgical techniques, and use of IUI and ART are covered to allow for optimal patient management. Recommendations are based on a strict process of evaluation of published literature as discussed in the Methodology section. This process is based on the PICO question approach (Problem/Patient/Population, Intervention/Indicator, Comparison, and Outcome) as described in the Methodology section. In this Guideline, the term “male” is used to refer to biological or genetic men.

Methodology

The Emergency Care Research Institute (ECRI) Evidence-based Practice Center team searched PubMed®, EMBASE® (Excerpta Medica), and Medline from January 2000 through May 2019. An experienced medical librarian developed an individual search strategy for each individual key question using medical subject headings terms and key words appropriate for each question’s PICO framework. When sufficient evidence existed, the body of evidence was assigned a strength rating of A (high), B (moderate), or C (low) for support of Strong, Moderate, or Conditional Recommendations. In the absence of sufficient evidence, additional information is provided as Clinical Principles and Expert Opinions. In 2023, the Male Infertility Guideline was updated through the AUA amendment process in which newly published literature is reviewed and integrated into previously published guidelines. ECRI’s medical research librarian conducted literature searches of EMBASE (Excerpta Medica)/Medline and PubMed (PreMedline) from May 30, 2019, through August 30, 2023, yielding 4,093 new abstracts. ECRI’s research analysts performed abstract screening and data extraction of 125 eligible study abstracts meeting inclusion criteria. There were 22 studies of interest that were included in the evidence base.

GUIDELINE STATEMENTS

Assessment

1. For initial infertility evaluation, clinicians should initiate concurrent assessment of both male and female partners. (*Expert Opinion*)
2. Clinicians should include a reproductive history during initial evaluation of the male for fertility. (*Clinical Principle*)
Clinicians should also include one or more semen analyses (SAs) during initial evaluation of the male. (*Strong Recommendation; Evidence Level: Grade B*)
3. Male reproductive experts should evaluate patients with a complete history and physical examination as well as other directed tests, when indicated by one or more abnormal semen parameters or presumed male infertility. (*Expert Opinion*)
4. In couples with failed assisted reproductive technology cycles or recurrent pregnancy losses (RPL) (two or more), clinicians should evaluate the male partner. (*Moderate Recommendation; Evidence Level: Grade C*)

Lifestyle Factors and Relationships Between Infertility and General Health

5. Clinicians should counsel infertile males or males with abnormal semen parameters on the health risks associated with abnormal sperm production. (*Moderate Recommendation; Evidence Level: Grade B*)
6. For infertile males with specific, identifiable causes of male infertility, clinicians should inform the patient of relevant, associated health conditions. (*Moderate Recommendation; Evidence Level: Grade B*)
7. Clinicians should advise couples with advanced paternal age (≥ 40) that there is an increased risk of adverse health outcomes for their offspring. (*Expert Opinion*)
8. Clinicians may discuss risk factors (i.e., lifestyle, medication usage, environmental exposures, occupational exposures) associated with male infertility, and counsel the patients that the current data on the majority of risk factors are limited. (*Conditional Recommendation; Evidence Level: Grade C*)

Diagnosis/Assessment/Evaluation

9. Clinicians should use the results from the semen analysis to guide management of the patient. In general, results are of greatest clinical significance when multiple abnormalities are present. (*Expert Opinion*)
10. Clinicians should obtain hormonal evaluation including follicle-stimulating hormone (FSH) and testosterone for infertile males with impaired libido, erectile dysfunction, oligozoospermia or azoospermia, atrophic testes, or evidence of hormonal abnormality on physical evaluation. (*Expert Opinion*)
11. Clinicians should initially evaluate azoospermic males with physical exam, semen volume, semen pH, and serum follicle-stimulating hormone levels to differentiate genital tract obstruction from impaired sperm production. (*Expert Opinion*)
12. Clinicians should recommend karyotype testing for males with primary infertility and azoospermia or sperm concentration < 5 million sperm/mL when accompanied by elevated follicle-stimulating hormone, testicular atrophy, or a diagnosis of impaired sperm production. (*Expert Opinion*)
13. Clinicians should recommend Y-chromosome microdeletion analysis for males with primary infertility and azoospermia or sperm concentration ≤ 1 million sperm/mL when accompanied by elevated follicle-stimulating hormone, testicular atrophy, or a diagnosis of impaired sperm production. (*Moderate Recommendation; Evidence Level: Grade B*)
14. Clinicians should recommend Cystic Fibrosis Transmembrane Conductance Regulator (*CFTR*) mutation carrier testing (including assessment of the 5T allele) in males with vasal agenesis or idiopathic obstructive azoospermia. (*Expert Opinion*)
15. For males who harbor a *CFTR* mutation or have absence of the vas deferens (unilateral or bilateral), clinicians should recommend genetic evaluation of the female partner. (*Expert Opinion*)
16. Clinicians should not recommend sperm deoxyribonucleic acid (DNA) fragmentation analysis in the initial evaluation of the infertile couple. (*Moderate Recommendation; Evidence Level: Grade C*)
17. In males with increased round cells on semen analysis (> 1 million/mL), clinicians should evaluate the patient further to differentiate white blood cells (pyospermia) from germ cells. (*Expert Opinion*)
18. In patients with pyospermia, clinicians should evaluate the patient for the presence of infection. (*Clinical Principle*)
19. Clinicians should not perform antisperm antibody (ASA) testing in the initial evaluation of male infertility. (*Expert Opinion*)
20. For couples with recurrent pregnancy loss, clinicians should evaluate the male partner with karyotype (*Expert Opinion*) and sperm DNA fragmentation. (*Moderate Recommendation; Evidence Level: Grade C*)
21. Clinicians should not routinely perform diagnostic testicular biopsy to differentiate between obstructive azoospermia and non-obstructive azoospermia (NOA). (*Expert Opinion*)

Imaging

22. Clinicians should not routinely perform scrotal ultrasound in the initial evaluation of the infertile male. (*Expert Opinion*)

23. Clinicians should not perform transrectal ultrasonography (TRUS) or pelvic magnetic resonance imaging (MRI) as part of the initial evaluation of the infertile male. Clinicians may recommend TRUS or pelvic MRI in males with semen analysis suggestive of ejaculatory duct obstruction (EDO) (i.e., acidic, azoospermic semen with volume <1.4mL, with normal serum T, palpable vas deferens). *(Expert Opinion)*
24. Clinicians should not routinely perform abdominal imaging for the sole indication of an isolated small or moderate right varicocele. *(Expert Opinion)*
25. Clinicians should recommend renal ultrasonography for patients with vasal agenesis to evaluate for renal abnormalities. *(Expert Opinion)*

Treatment

VARICOCELE REPAIR/VARICOCELECTOMY

26. Clinicians should consider surgical varicocelectomy in males attempting to conceive who have palpable varicocele(s), infertility, and abnormal semen parameters, except for azoospermic males. *(Moderate Recommendation; Evidence Level: Grade B)*
27. Clinicians should not recommend varicocelectomy for males with non-palpable varicoceles detected solely by imaging. *(Strong Recommendation; Evidence Level: Grade C)*
28. For males with clinical varicocele and non-obstructive azoospermia, clinicians should inform couples of the absence of definitive evidence supporting varicocele repair prior to surgical sperm retrieval with assisted reproductive technologies. *(Expert Opinion)*

SPERM RETRIEVAL

29. For males with non-obstructive azoospermia undergoing sperm retrieval, clinicians should perform a microdissection testicular sperm extraction (micro-TESE). *(Moderate Recommendation; Evidence Level: Grade C)*
30. In males undergoing surgical sperm retrieval by a clinician, intracytoplasmic sperm injection may be performed with fresh or cryopreserved sperm. *(Conditional Recommendation; Evidence Level: Grade C)*
31. In males with azoospermia due to obstruction undergoing surgical sperm retrieval, clinicians may extract sperm from either the testis or the epididymis. *(Conditional Recommendation; Evidence Level: Grade C)*
32. Clinicians may consider the utilization of testicular sperm in nonazoospermic males with an elevated sperm DNA Fragmentation Index (DFI). *(Clinical Principle)*
33. For males with aspermia, clinicians may perform surgical sperm extraction or induced ejaculation (sympathomimetics, vibratory stimulation or electroejaculation) depending on the patient's condition and clinician's experience. *(Expert Opinion)*
34. Clinicians may treat infertility associated with retrograde ejaculation (RE) with sympathomimetics (with or without alkalization and/or urethral catheterization), induced ejaculation, or surgical sperm retrieval. *(Expert Opinion)*

OBSTRUCTIVE AZOOSPERMIA, INCLUDING POST-VASECTOMY INFERTILITY

35. Clinicians should counsel couples desiring conception after vasectomy that surgical reconstruction, surgical sperm retrieval, or both reconstruction and simultaneous sperm retrieval for cryopreservation are viable options. *(Moderate Recommendation; Evidence Level: Grade C)*
36. Clinicians should counsel males with vasal or epididymal obstructive azoospermia that microsurgical reconstruction may be successful in returning sperm to the ejaculate. *(Expert Opinion)*
37. For infertile males with ejaculatory duct obstruction, clinicians may consider transurethral resection of ejaculatory ducts (TURED) and/or surgical sperm extraction. *(Expert Opinion)*

MEDICAL AND NUTRACEUTICAL INTERVENTIONS FOR FERTILITY

38. Clinicians may manage male infertility with assisted reproductive technology. *(Expert Opinion)*
39. Clinicians may advise an infertile couple with a low total motile sperm count on repeated semen analyses that intrauterine insemination success rates may be reduced, and treatment with assisted reproductive technology (in vitro fertilization/intracytoplasmic sperm injection) may be considered. *(Expert Opinion)*

40. In a patient presenting with hypogonadotropic hypogonadism (HH), clinicians should evaluate the patient to determine the etiology of the disorder and treat based on diagnosis. (*Clinical Principle*)
41. Clinicians may use aromatase inhibitors (AIs), human chorionic gonadotropin (hCG), selective estrogen receptor modulators (SERMs), or a combination thereof for infertile males with low serum testosterone. (*Conditional Recommendation; Evidence Level: Grade C*)
42. For the male interested in current or future fertility, clinicians should not prescribe exogenous testosterone therapy. (*Clinical Principle*)
43. For the infertile male with hyperprolactinemia, clinicians should evaluate the patient for the etiology and treat accordingly. (*Expert Opinion*)
44. Clinicians should inform the male with idiopathic infertility that the use of selective estrogen receptor modulators has limited benefits relative to results of assisted reproductive technology. (*Expert Opinion*)
45. Clinicians should counsel patients that the benefits of supplements (e.g., antioxidants, vitamins) are of questionable clinical utility in treating male infertility. Existing data are inadequate to provide recommendation for specific agents to use for this purpose. (*Moderate Recommendation; Evidence Level: Grade B*)
46. For males with idiopathic infertility, clinicians may consider treatment using a follicle-stimulating hormone analogue with the aim of improving sperm concentration, pregnancy rate, and live birth rate. (*Conditional Recommendation; Evidence Level: Grade B*)
47. In patients with non-obstructive azoospermia, clinicians may inform the patient of the limited data supporting pharmacologic manipulation with selective estrogen receptor modulators, aromatase inhibitors, and gonadotropins prior to surgical intervention. (*Conditional Recommendation; Evidence Level: Grade C*)

GONADOTOXIC THERAPIES AND FERTILITY PRESERVATION

48. Clinicians should discuss the effects of gonadotoxic therapies and other cancer treatments on sperm production with patients prior to commencement of therapy. (*Moderate Recommendation; Evidence Level: Grade C*)
49. Clinicians should inform patients undergoing chemotherapy and/or radiation therapy to avoid initiating a pregnancy for a period of at least 12 months after completion of treatment. (*Expert Opinion*)
50. Clinicians should encourage males to bank sperm, preferably multiple specimens when possible, prior to commencement of gonadotoxic therapy or other cancer treatment that may affect fertility in males. (*Expert Opinion*)
51. Clinicians may inform patients that a semen analysis should be performed at least 12 months (and preferably 24 months) after completion of gonadotoxic therapies. (*Conditional Recommendation; Evidence Level: Grade C*)
52. Clinicians should inform patients undergoing a retroperitoneal lymph node dissection (RPLND) of the risk of aspermia or retrograde ejaculation. (*Clinical Principle*)
53. Clinicians should obtain a post-orgasmic urinalysis for males with aspermia after retroperitoneal lymph node dissection and reduced volume ejaculate who are interested in fertility. (*Clinical Principle*)
54. Clinicians should inform males seeking paternity who are persistently azoospermic after gonadotoxic therapies that microdissection testicular sperm extraction is a treatment option. (*Strong Recommendation; Evidence Level: Grade B*)

INTRODUCTION

The Diagnosis and Treatment of the Male Factor Couple

Approximately 15% of couples experience infertility, which is defined as the inability to conceive a pregnancy within 12 months when the female partner is <35 years old and within 6 months when the female partner is >35 years old. A male factor is solely responsible in about 20% of infertile couples and contributory in another 30% to 40%.¹ Despite these estimates, the true prevalence of male infertility is not clearly defined due to multiple factors including variations in definitions of infertility, differences in sources of data, and the populations studied.² Male factor infertility may be explained by an abnormal SA or by other sperm function defects, in the setting of a normal SA as well as functional m defects. This document offers guidance for the optimal diagnostic evaluation and management of the male partner o. infertile couple.

Male infertility can be due to a variety of conditions. Some of these conditions are identifiable and reversible, such as ductal obstruction and HH. Other conditions are identifiable and treatable but not reversible, such as bilateral testicular atrophy secondary to viral orchitis. Identification of the etiology of an abnormal SA is not possible in approximately 30% of males in which case this condition is termed idiopathic male infertility.³ When the reason for infertility is not clear with a normal SA and partner evaluation the infertility is termed unexplained, which is found in up to approximately 25% of couples.³ In some instances, patients with normal SAs have sperm that do not function in a manner necessary for fertility.

The overall goal of the male evaluation is to identify conditions that may affect management or health of the patient or their offspring. Identification and treatment of reversible conditions may improve the male's fertility and allow for conception through intercourse or through techniques, such as IUI or IVF, when those approaches would otherwise not be possible. Even azoospermic patients may have some degree of active sperm production within the testes or could have sperm production induced with treatment. Identification of conditions for which there is no treatment will spare couples the distress of attempting ineffective therapies and allow them to consider options, such as donor sperm or adoption, if appropriate. Male infertility is associated with other comorbidities including increased mortality, while advanced paternal age is associated with some adverse outcomes in offspring. In addition, male infertility may occasionally be the presenting manifestation of an underlying life-threatening condition.⁴ Failure to identify diseases such as testicular cancer or pituitary tumors may have serious consequences, including, in rare cases, death. Detection of certain genetic causes of male infertility allows couples to be informed about the potential to transmit genetic abnormalities that may affect the health of offspring and seek genetic counseling when appropriate. Thus, an appropriate male evaluation may allow the couple to better understand the basis and implications of their infertility.

In summary, the specific goals of the evaluation of the infertile male are to identify the following:

- potentially correctable conditions;
- irreversible conditions that are amenable to IUI and ART using the sperm of the male partner;
- irreversible conditions that are not amenable to the above, and for which donor insemination or adoption are possible options;
- life- or health-threatening conditions that may underlie the infertility or associated medical comorbidities that require medical attention; and
- genetic abnormalities or lifestyle and age factors that may affect the health of the male patient or of offspring particularly if ART are to be employed.

Definitions of Infertility and Treatment Success

A wide variety of professional and international health organizations have defined infertility in general and male infertility, specifically. Since the condition of infertility reflects the outcome of a couple's attempt to achieve a pregnancy, the most common definition of infertility is "a disease of the reproductive system defined by the failure to achieve a clinical pregnancy after 12 months or more of regular unprotected sexual intercourse".⁵ The condition of infertility is categorized as a disease by the World Health Organization (WHO), the American Medical Association (AMA), and the American Society for Reproductive Medicine (ASRM).⁶ Evaluation for infertility is also guided by female age and other factors, such as an abnormal male reproductive history (e.g., history of cryptorchidism, chemotherapy, pelvic/retroperitoneal surgery, other conditions that have been associated with male infertility). When such factors are present, male evaluation is indicated regardless of prior attempts to conceive. Infertility should be evaluated after 6 months of attempted conception when the female partner is 35 years of age or older.

Male infertility is typically diagnosed by one or more factors that may include abnormal semen quality or sperm functional parameters; anatomical, endocrine, genetic, functional, or immunological abnormalities of the male reproductive system (including chronic illness); or sexual conditions (e.g., erectile dysfunction) incompatible with the ability to deposit semen in the vagina. Primary male infertility refers to males who have never initiated a clinical pregnancy and meets the criteria of being classified as infertile, whereas secondary infertility refers to a couple where the male is unable to initiate a clinical pregnancy, but who had previously initiated a clinical pregnancy (with the same or different sexual partner). Some conditions may be more common in primary or secondary infertility. Evaluation of males with secondary infertility should include a focus on conditions or exposures that have developed or occurred after initiation of the earlier pregnancy(ies).

Assessment of tests and treatments for the male is challenging due to inconsistent endpoints and the observation that many of these endpoints are dependent upon and measured from the female partner. Ideally, the endpoint for fertility trials should be "live birth (defined as any delivery of a live infant after 20 weeks of gestation) or cumulative live birth, defined as the live birth per women over a defined time period (or number of treatment cycles)." This definition was

provided by the modified Consolidated Standards of Reporting Trials for Fertility, Improving the Reporting of Clinical Trials of Infertility Treatments.⁷ However, due to the variety of confounding variables present in the female, it is difficult to control for many of the most important variables and still include sufficient male subjects in a clinical trial for pregnancy or birth to be a viable outcome measure.

To address this challenge, the majority of clinical trials addressing male fertility and infertility utilize surrogate outcome metrics, the most common being the SA. However, the high variability of SA parameters make them difficult to use in the determination of interventions for male reproduction.⁵ Other outcome metrics with similar challenges include other types of sperm tests and ART outcomes such as fertilization, implantation, and pregnancy loss rates. All attempts to measure some aspect of sperm function lessens the confounder effect of a maternal outcome, yet all are also subject to their own limitations. Common terms used in semen analysis can be found in **Table 1**.

Table 1: Common Terms in Semen Analysis*

Term	Definition
Aspermia	Complete absence of semen in ejaculate, indicating the absence of seminal fluid production or blockage in the reproductive tract.
Azoospermia	Absence of spermatozoa in the semen, typically resulting from a blockage in the reproductive tract (obstructive azoospermia) or dysfunction in sperm production (non-obstructive azoospermia).
Oligozoospermia	Low sperm concentration in the semen.
Asthenozoospermia	Reduced sperm motility, where a significant portion of spermatozoa display sluggish or abnormal movement, impacting fertility.
Teratozoospermia	Abnormal sperm morphology, characterized by a high percentage of spermatozoa with morphological defects, potentially affecting fertility.
Normozoospermia	Normal semen parameters including sperm concentration, motility, morphology, and volume, indicating optimal fertility potential.
Retrograde Ejaculation	Condition where semen flows backward into the bladder instead of exiting through the urethra during ejaculation, leading to reduced fertility.
* This table provides concise definitions for common terms used in semen analysis, facilitating understanding for researchers, clinicians, and individuals seeking information about male fertility assessment.^{8,9}	

Epidemiology

Most couples achieve a pregnancy in the first 3 to 6 months of attempted conception, with 75% of couples achieving a pregnancy after 6 months of trying.¹⁰⁻¹³ In general, after one year of attempting to conceive, approximately 85% of couples will have achieved a pregnancy. After two full years of attempting to conceive, this statistic is increased to over 90% of couples.

Age of the female partner is the single most important factor when predicting the chances of conception for a couple. Fertility decreases by almost 50% in women in their late 30's compared to women in their 20's. In women under 35 years of age, infertility is considered present after 12 months of attempting to conceive. This duration is shortened to 6 months in women 35 years of age or older.^{14,15}

The etiologic causes of infertility include both female and male factors. For women, these factors include ovulatory dysfunction, tubal factor, endometriosis, and uterine factors. For the woman, ovarian reserve is helpful in predicting her response to medications, but this is not an absolute predictor of fertility. In up to 60% of couples, a male factor is found as part of the etiology of the infertility.¹⁶ In addition, approximately 25% of couples will have unexplained infertility.

RPL is a disease that is distinct from infertility and is defined as two or more failed pregnancies.⁶ The workup of RPL yields an etiology in only approximately 50% of couples as most pregnancy losses are related to abnormalities within the fetus itself. The risk of pregnancy loss after two losses is at least 25% depending on the age of the woman. After three consecutive losses, this risk increases to almost 50%. Etiologic causes of recurrent pregnancy losses include genetic causes (e.g., chromosomal translocations), anatomic abnormalities of the female uterus (e.g., septum, submucosal fibroids, adhesions), infections, hematologic and immunologic disorders of the female partner, female partner endocrine issues (e.g., thyroid and diabetes), and male factor issues.¹⁷⁻¹⁹ In general, for males, the common identified etiologic issues include karyotypic abnormalities and sperm DNA fragmentation.

Methodology

PANEL FORMATION

The Male Infertility Panel was created in 2017 by the American Urological Association Education and Research, Inc. (AUAER) and ASRM. The AUA Practice Guidelines Committee (PGC) selected the Panel Chairs, who in turn appointed the additional panel members based on specific expertise in this area. The Panel included specialties from urology, andrology, endocrinology, and obstetrics & gynecology. There was also a patient advocate representative from RESOLVE: The National Infertility Association. The Male Infertility Amendment Panel was created in 2023 by the AUA to review new literature and provide updates herein. The Panel received no remuneration for their work.

SEARCHES AND ARTICLE SELECTION

The Emergency Care Research Institute (ECRI) Evidence-based Practice Center team searched PubMed®, Embase®, and Medline from January 2000 through May 2019. An experienced medical librarian developed an individual search strategy for each individual key question using medical subject headings terms and key words appropriate for each question's PICO framework. Search strategies were reviewed by one of the project methodologists. The evidence review team also reviewed relevant systematic reviews and references provided by the Panel to identify articles that may have been missed by the database searches. In 2023, the Male Infertility Guideline was updated through the AUA amendment process in which newly published literature is reviewed and integrated into previously published guidelines. ECRI's medical research librarian conducted literature searches of EMBASE (Excerpta Medica)/Medline and PubMed (PreMedline) from May 30, 2019, through August 30, 2023, yielding 4,093 new abstracts. ECRI's research analysts performed abstract screening and data extraction of 125 eligible study abstracts meeting inclusion criteria.

DATA ABSTRACTIONS

Study selection was based on predefined eligibility criteria for the patient populations, interventions, outcomes, and study designs of interest. Two reviewers independently screened abstracts and full text for inclusion. Conflicts between reviewers regarding eligibility of a given study were resolved through consensus.

Reviewers extracted information on study characteristics, participants, interventions, and outcomes. One reviewer completed data abstraction for each included study.

Members of the AUA Male Infertility Guideline Amendment Panel met with ECRI research analysts in July 2023 to review the recommendation statements from the 2020 Guideline. The Panel identified clinical recommendations for which they asked ECRI to perform an updated search to assess newly retrieved abstracts to inform possible updating of the recommendations. ECRI research analysts mapped the clinical recommendations of interest to the key questions in the systematic review to which they pertained.

RISK OF BIAS ASSESSMENT

One reviewer independently assessed risk of bias (ROB) for individual studies. The Cochrane Collaboration's tool was used for assessing the risk of bias of randomized controlled trials (RCTs).²⁰ For non-randomized studies of treatment interventions, the reviewers used appropriate items from the Cochrane Risk of Bias Assessment Tool for Non-Randomized Studies of Interventions (ACROBAT-NRSI). For diagnostic studies, reviewers used the quality assessment tool for diagnostic accuracy studies (QUADAS -2).²¹ Single-arm studies were assessed by the following domains: prospective or retrospective design, consecutive/non-consecutive enrollment, incomplete outcome data, selective outcome reporting, and any other potential sources of bias. For systematic reviews, ROB was assigned based on the study authors' quality assessment of the individual studies included in the review. If such an assessment was not provided, ECRI analysts assigned a ROB rating based on the author description of the selected literature base and the designs of the included studies. The evidence review team graded strength of evidence on outcomes by adapting the AUA's three predefined levels of strength of evidence.

DETERMINATION OF EVIDENCE STRENGTH

The AUA employs a 3-tiered strength of evidence system to underpin evidence-based guideline statements. **Table 2** summarizes the GRADE categories, definitions, and how these categories translate to the AUA strength of evidence categories. In short, high certainty by GRADE translates to AUA A-category strength of evidence, moderate to B, and both low and very low to C.

The categorization of evidence strength is conceptually distinct from the quality of individual studies. Evidence strength refers to the body of evidence available for a particular question and includes not only the quality of individual studies; consideration of study design; consistency of findings across studies; adequacy of sample sizes; and generalizability of populations, settings, and interventions for the purposes of the Guideline. The AUA categorizes body of evidence strength as Grade A (well-conducted and highly-generalizable RCTs or exceptionally strong observational studies with consistent

findings), Grade B (RCTs with some weaknesses of procedure or generalizability or moderately strong observational studies with consistent findings), or Grade C (RCTs with serious deficiencies of procedure or generalizability or extremely small sample sizes or observational studies that are inconsistent, have small sample sizes, or have other problems that potentially confound interpretation of data). By definition, Grade A evidence has a high level of certainty, Grade B evidence has a moderate level of certainty, and Grade C evidence has a low level of certainty.²²

Table 2: Strength of Evidence Definitions

AUA Strength of Evidence Category	GRADE Certainty Rating	Definition
A	High	• Very confident that the true effect lies close to that of the estimate of the effect
B	Moderate	• Moderately confident in the effect estimate • The true effect is likely to be close to the estimate of the effect, but there is a possibility that it is substantially different
C	Low	• Confidence in the effect estimate is limited • The true effect may be substantially different from the estimate of the effect
	Very Low	• Very little confidence in the effect estimate • The true effect is likely to be substantially different from the estimate of effect

AUA NOMENCLATURE: LINKING STATEMENT TYPE TO EVIDENCE STRENGTH

The AUA nomenclature system explicitly links statement type to body of evidence strength, level of certainty, magnitude of benefit or risk/burdens, and the Panel's judgment regarding the balance between benefits and risks/burdens (Table 3).

Strong Recommendations are directive statements that an action should (benefits outweigh risks/burdens) or should not (risks/burdens outweigh benefits) be undertaken because net benefit or net harm is substantial. **Moderate**

Recommendations are directive statements that an action should (benefits outweigh risks/burdens) or should not (risks/burdens outweigh benefits) be undertaken because net benefit or net harm is moderate. **Conditional**

Recommendations are non-directive statements used when the evidence indicates there is no apparent net benefit or harm or when the balance between benefits and risks/burden is unclear. All three statement types may be supported by any body of evidence strength grade. Body of evidence strength Grade A in support of a Strong or Moderate Recommendation indicates the statement can be applied to most patients in most circumstances and that future research is *unlikely to change confidence*. Body of evidence strength Grade B in support of a Strong or Moderate Recommendation indicates the statement can be applied to most patients in most circumstances, but better evidence *could change confidence*. Body of evidence strength Grade C in support of a Strong or Moderate Recommendation indicates the statement can be applied to most patients in most circumstances, but better evidence is *likely to change confidence*. Conditional Recommendations can also be supported by any evidence strength. When body of evidence strength is Grade A, the statement indicates benefits and risks/burdens appear balanced, the best action depends on patient circumstances, and future research is *unlikely to change confidence*. When body of evidence strength Grade B is used, benefits and risks/burdens appear balanced, the best action also depends on individual patient circumstances, and better evidence *could change confidence*. When body of evidence strength Grade C is used, there is uncertainty regarding the balance between benefits and risks/burdens, alternative strategies may be equally reasonable, and better evidence is *likely to change confidence*.

Where gaps in the evidence existed, Clinical Principles or Expert Opinions are provided via consensus of the Panel. A **Clinical Principle** is a statement about a component of clinical care widely agreed upon by urologists or other clinicians for which there may or may not be evidence in the medical literature. **Expert Opinion** refers to a statement based on members' clinical training, experience, knowledge, and judgment for which there may or may not be evidence in the medical literature.

Table 3: AUA Nomenclature Linking Statement Type to Level of Certainty, Magnitude of Benefit or Risk/Burden, and Body of Evidence Strength

Evidence Grade	Evidence Strength A (High Certainty)	Evidence Strength B (Moderate Certainty)	Evidence Strength C (Low Certainty)
Strong Recommendation (Net benefit or harm substantial)	-Benefits > Risks/Burdens (or vice versa) -Net benefit (or net harm) is substantial -Applies to most patients in most circumstances and future research is unlikely to change confidence	-Benefits > Risks/Burdens (or vice versa) -Net benefit (or net harm) is substantial -Applies to most patients in most circumstances but better evidence could change confidence	-Benefits > Risks/Burdens (or vice versa) -Net benefit (or net harm) appears substantial -Applies to most patients in most circumstances but better evidence is likely to change confidence (rarely used to support a Strong Recommendation)
Moderate Recommendation (Net benefit or harm moderate)	-Benefits > Risks/Burdens (or vice versa) -Net benefit (or net harm) is moderate -Applies to most patients in most circumstances and future research is unlikely to change confidence	-Benefits > Risks/Burdens (or vice versa) -Net benefit (or net harm) is moderate -Applies to most patients in most circumstances but better evidence could change confidence	-Benefits > Risks/Burdens (or vice versa) -Net benefit (or net harm) appears moderate -Applies to most patients in most circumstances but better evidence is likely to change confidence
Conditional Recommendation (Net benefit or harm comparable to other options)	-Benefits=Risks/Burdens -Best action depends on individual patient circumstances -Future Research is unlikely to change confidence	-Benefits=Risks/Burdens -Best action appears to depend on individual patient circumstances -Better evidence could change confidence	-Balance between Benefits & Risks/Burdens unclear -Net benefit (or net harm) comparable to other options -Alternative strategies may be equally reasonable -Better evidence likely to change confidence
Clinical Principle	a statement about a component of clinical care that is widely agreed upon by urologists or other clinicians for which there may or may not be evidence in the medical literature		
Expert Opinion	a statement, achieved by consensus of the Panel, that is based on members' clinical training, experience, knowledge, and judgment for which there may or may not be evidence in the medical literature		

PEER REVIEW AND DOCUMENT APPROVAL

An integral part of the Guideline development process at the AUA is external peer review. The AUA conducted a thorough peer review process to ensure that the document was reviewed by experts in the diagnosis and treatment of male infertility. In addition to reviewers from the AUA PGC, Science and Quality Council (SQC), and Board of Directors (BOD), the document was reviewed by representatives from ASRM, as well as external content experts. Additionally, a call for reviewers was placed on the AUA website from January 8-15, 2020 to allow any further interested parties to request a copy of the document for review. The Guideline was also sent to the Urology Care Foundation to open the document further to the patient perspective. The draft Guideline document was distributed to 114 peer reviewers. All peer review comments were blinded and sent to the Panel for review. In total, 49 reviewers provided comments, including 24 external reviewers. At the end of the peer review process, a total of 997 comments were received. Following comment discussion, the Panel revised the draft as needed. Once finalized, the Guideline was submitted for approval to the AUA PGC, SQC, and BOD for final approval. The document was also approved by the ASRM CEO Ricardo Azziz, MD, MPH, MBA, on behalf of the Board and advised by the Practice Committee.

In 2024, as a part of the amendment process, the AUA conducted a thorough peer review process. A call for peer reviewers was posted on March 5th, 2024 and the draft Guideline document was distributed to 111 peer reviewers, 51 of which submitted comments. The Amendment Panel reviewed and discussed all submitted comments and revised the draft as needed. Once finalized, the Guideline was submitted for approval to the PGC and SQC as well as representatives from ASRM. It was then submitted to AUA BODs for final approval.

GUIDELINE STATEMENTS

Assessment

GUIDELINE STATEMENT 1

For initial infertility evaluation, clinicians should initiate concurrent assessment of both male and female partners. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 2

Clinicians should include a reproductive history during initial evaluation of the male for fertility. (*Clinical Principle*)
Clinicians should also include one or more semen analyses during initial evaluation of the male. (*Strong Recommendation; Evidence Level: Grade B*)

Discussion

GUIDELINE STATEMENT 3

Male reproductive experts should evaluate patients with a complete history and physical examination as well as other directed tests, when indicated by one or more abnormal semen parameters or presumed male infertility. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 4

In couples with failed assisted reproductive technology cycles or recurrent pregnancy losses (two or more), clinicians should evaluate the male partner. (*Moderate Recommendation; Evidence Level: Grade C*)

Discussion

Lifestyle Factors and Relationships Between Infertility and General Health

GUIDELINE STATEMENT 5

Clinicians should counsel infertile males or males with abnormal semen parameters on the health risks associated with abnormal sperm production. (*Moderate Recommendation; Evidence Level: Grade B*)

Discussion

GUIDELINE STATEMENT 6

For infertile males with specific, identifiable causes of male infertility, clinicians should inform the patient of relevant, associated health conditions. (*Moderate Recommendation; Evidence Level: Grade B*)

Discussion

GUIDELINE STATEMENT 7

Clinicians should advise couples with advanced paternal age (≥ 40) that there is an increased risk of adverse health outcomes for their offspring. (*Expert Opinion*)

GUIDELINE STATEMENT 8

Clinicians may discuss risk factors (i.e., lifestyle, medication usage, environmental exposures, occupational exposures) associated with male infertility, and counsel the patients that the current data on the majority of risk factors are limited. **(Conditional Recommendation; Evidence Level: Grade C)**

Diagnosis and Evaluation

GUIDELINE STATEMENT 9

Clinicians should use the results from the semen analysis to guide management of the patient. In general, results are of greatest clinical significance when multiple abnormalities are present. **(Expert Opinion)**

GUIDELINE STATEMENT 10

Clinicians should obtain hormonal evaluation including follicle-stimulating hormone and testosterone for infertile males with impaired libido, erectile dysfunction, oligozoospermia or azoospermia, atrophic testes, or evidence of hormonal abnormality on physical evaluation. **(Expert Opinion)**

GUIDELINE STATEMENT 11

Clinicians should initially evaluate azoospermic males with physical exam, semen volume, semen pH, and serum follicle-stimulating hormone levels to differentiate genital tract obstruction from impaired sperm production. **(Expert Opinion)**

GUIDELINE STATEMENT 12

Clinicians should recommend karyotype testing for males with primary infertility and azoospermia or sperm concentration <5 million sperm/mL when accompanied by elevated follicle-stimulating hormone, testicular atrophy, or a diagnosis of impaired sperm production. **(Expert Opinion)**

GUIDELINE STATEMENT 13

Clinicians should recommend Y-chromosome microdeletion analysis for males with primary infertility and azoospermia or sperm concentration ≤ 1 million sperm/mL when accompanied by elevated follicle-stimulating hormone, testicular atrophy, or a diagnosis of impaired sperm production. **(Moderate Recommendation; Evidence Level: Grade B)**

GUIDELINE STATEMENT 14

Clinicians should recommend Cystic Fibrosis Transmembrane Conductance Regulator (**CFTR**) mutation carrier testing (including assessment of the 5-thymidine [5T] allele) in males with vasal agenesis or idiopathic obstructive azoospermia. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 15

For males who harbor a **CFTR** mutation or have absence of the vas deferens (unilateral or bilateral), clinicians should recommend genetic evaluation of the female partner. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 16

Clinicians should not recommend sperm DNA fragmentation analysis in the initial evaluation of the infertile couple. (*Moderate Recommendation; Evidence Level: Grade C*)

Discussion

GUIDELINE STATEMENT 17

In males with increased round cells on semen analysis (>1 million/mL), clinicians should evaluate the patient further to differentiate white blood cells (pyospermia) from germ cells. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 18

In patients with pyospermia, clinicians should evaluate the patient for the presence of infection. (*Clinical Principle*)

Discussion

GUIDELINE STATEMENT 19

Clinicians should not perform antisperm antibody (ASA) testing in the initial evaluation of male infertility. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 20

For couples with recurrent pregnancy loss (RPL), clinicians should evaluate the male partner with karyotype (*Expert Opinion*) and sperm DNA fragmentation. (*Moderate Recommendation; Evidence Level: Grade C*)

Discussion

GUIDELINE STATEMENT 21

Clinicians should not routinely perform diagnostic testicular biopsy to differentiate between obstructive azoospermia non-obstructive azoospermia (NOA). (*Expert Opinion*)

Imaging

GUIDELINE STATEMENT 22

Clinicians should not routinely perform scrotal ultrasound in the initial evaluation of the infertile male. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 23

Clinicians should not perform transrectal ultrasonography (TRUS) or pelvic MRI as part of the initial evaluation of the infertile male. Clinicians may recommend TRUS or pelvic MRI in males with SA suggestive of ejaculatory duct obstruction (EDO) (i.e., acidic, azoospermic semen with volume <1.4 mL, with normal serum T, palpable vas deferens). (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 24

Clinicians should not routinely perform abdominal imaging for the sole indication of an isolated small or moderate right varicocele. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 25

Clinicians should recommend renal ultrasonography for patients with vasal agenesis to evaluate for renal abnormalities. (*Expert Opinion*)

Discussion

Treatment

VARICOCELE REPAIR/VARICOCELECTOMY

GUIDELINE STATEMENT 26

Clinicians should consider surgical varicocelectomy in males attempting to conceive who have palpable varicocele(s), infertility, and abnormal semen parameters, except for azoospermic males. (*Moderate Recommendation; Evidence Level: Grade B*)

Discussion

GUIDELINE STATEMENT 27

Clinicians should not recommend varicocelectomy for males with non-palpable varicoceles detected solely by imaging. (*Strong Recommendation; Evidence Level: Grade C*)

Discussion

GUIDELINE STATEMENT 28

For males with clinical varicocele and non-obstructive azoospermia (NOA), clinicians should inform couples of the absence of definitive evidence supporting varicocele repair prior to surgical sperm retrieval with assisted reproductive technologies. (*Expert Opinion*)

Discussion

SPERM RETRIEVAL

GUIDELINE STATEMENT 29

For males with non-obstructive azoospermia undergoing sperm retrieval, clinicians should perform a microdissection testicular sperm extraction. (*Moderate Recommendation; Evidence Level: Grade C*)

Discussion

GUIDELINE STATEMENT 30

In males undergoing surgical sperm retrieval by a clinician, intracytoplasmic sperm injection may be performed with fresh or cryopreserved sperm. (*Conditional Recommendation; Evidence Level: Grade C*)

Discussion

GUIDELINE STATEMENT 31

In males with azoospermia due to obstruction undergoing surgical sperm retrieval, clinicians may extract sperm from either the testis or the epididymis. (*Conditional Recommendation; Evidence Level: Grade C*)

Discussion

GUIDELINE STATEMENT 32

Clinicians may consider the utilization of testicular sperm in nonazoospermic males with an elevated sperm DNA Fragmentation Index. (*Clinical Principle*)

Discussion

GUIDELINE STATEMENT 33

For males with aspermia, clinicians may perform surgical sperm extraction or induced ejaculation (sympathomimetics, vibratory stimulation or electroejaculation) depending on the patient's condition and clinician's experience. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 34

Clinicians may treat infertility associated with retrograde ejaculation with sympathomimetics (with or without alkalization and/or urethral catheterization), induced ejaculation, or surgical sperm retrieval. (*Expert Opinion*)

Discussion

OBSTRUCTIVE AZOOSPERMIA, INCLUDING POST-VASECTOMY INFERTILITY

GUIDELINE STATEMENT 35

Clinicians should counsel couples desiring conception after vasectomy that surgical reconstruction, surgical sperm retrieval, or both reconstruction and simultaneous sperm retrieval for cryopreservation are viable options. (*Moderate Recommendation; Evidence Level: Grade C*)

Discussion

GUIDELINE STATEMENT 36

Clinicians should counsel males with vasal or epididymal obstructive azoospermia that microsurgical reconstruction may be successful in returning sperm to the ejaculate. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 37

For infertile males with ejaculatory duct obstruction, clinicians may consider transurethral resection of ejaculatory ducts (TURED) and/or surgical sperm extraction. (*Expert Opinion*)

Discussion

MEDICAL AND NUTRACEUTICAL INTERVENTIONS FOR FERTILITY

GUIDELINE STATEMENT 38

Clinicians may manage male infertility with assisted reproductive technologies. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 39

Clinicians may advise an infertile couple with a low total motile sperm count on repeated semen analyses that intrauterine insemination success rates may be reduced, and treatment with assisted reproductive technologies (in vitro fertilization with intracytoplasmic sperm injection) may be considered. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 40

In a patient presenting with hypogonadotropic hypogonadism, clinicians should evaluate the patient to determine the etiology of the disorder and treat based on diagnosis. (*Clinical Principle*)

Discussion

GUIDELINE STATEMENT 41

Clinicians may use aromatase inhibitors, human chorionic gonadotropin, selective estrogen receptor modulators, or a combination thereof for infertile males with low serum testosterone. (*Conditional Recommendation; Evidence Level: Grade C*)

Discussion

GUIDELINE STATEMENT 42

For the male interested in current or future fertility, clinicians should not prescribe exogenous testosterone therapy. **(Clinical Principle)**

Discussion

GUIDELINE STATEMENT 43

For the infertile male with hyperprolactinemia, clinicians should evaluate the patient for the etiology and treat accordingly. **(Expert Opinion)**

Discussion

GUIDELINE STATEMENT 44

Clinicians should inform the male with idiopathic infertility that the use of selective estrogen receptor modulators has limited benefits relative to results of assisted reproductive technologies. **(Expert Opinion)**

Discussion

GUIDELINE STATEMENT 45

Clinicians should counsel patients that the benefits of supplements (e.g., antioxidants, vitamins) are of questionable clinical utility in treating male infertility. Existing data are inadequate to provide recommendation for specific agents to use for this purpose. **(Moderate Recommendation; Evidence Level: Grade B)**

Discussion

GUIDELINE STATEMENT 46

For males with idiopathic infertility, clinicians may consider treatment using a follicle-stimulating hormone analogue with the aim of improving sperm concentration, pregnancy rate, and live birth rate. **(Conditional Recommendation; Evidence Level: Grade B)**

Discussion

GUIDELINE STATEMENT 47

In patients with non-obstructive azoospermia, clinicians may inform the patient of the limited data supporting pharmacologic manipulation with selective estrogen receptor modulators, aromatase inhibitors, and gonadotropins prior to surgical intervention. **(Conditional Recommendation; Evidence Level: Grade C)**

Discussion

GONADOTOXIC THERAPIES AND FERTILITY PRESERVATION

GUIDELINE STATEMENT 48

Clinicians should discuss the effects of gonadotoxic therapies and other cancer treatments on sperm production with patients prior to commencement of therapy. **(Moderate Recommendation; Evidence Level: Grade C)**

GUIDELINE STATEMENT 49

Clinicians should inform patients undergoing chemotherapy and/or radiation therapy to avoid initiating a pregnancy for a period of at least 12 months after completion of treatment. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 50

Clinicians should encourage males to bank sperm, preferably multiple specimens when possible, prior to commencement of gonadotoxic therapy or other cancer treatment that may affect fertility in males. (*Expert Opinion*)

Discussion

GUIDELINE STATEMENT 51

Clinicians may inform patients that a semen analysis should be performed at least 12 months (and preferably 24 months) after completion of gonadotoxic therapies. (*Conditional Recommendation; Evidence Level: Grade C*)

Discussion

GUIDELINE STATEMENT 52

Clinicians should inform patients undergoing a retroperitoneal lymph node dissection of the risk of aspermia or retrograde ejaculation. (*Clinical Principle*)

Discussion

GUIDELINE STATEMENT 53

Clinicians should obtain a post-orgasmic urinalysis for males with aspermia after retroperitoneal lymph node dissection and reduced volume ejaculate who are interested in fertility. (*Clinical Principle*)

Discussion

GUIDELINE STATEMENT 54

Clinicians should inform males seeking paternity who are persistently azoospermic after gonadotoxic therapies that microdissection testicular sperm extraction is a treatment option. (*Strong Recommendation; Evidence Level: Grade B*)

Discussion

FUTURE DIRECTIONS

Newer research techniques, such as next generation sequencing (whole exome and whole genome sequencing) and “-omic” technologies have been applied to better identify underlying defects that may explain infertility in males. As the mechanisms of action of these genetic, genomic, epigenetic, transcriptomic, proteomic, metabolomic defects are defined, we will have further defined the etiologies of the majority of causes of male infertility. For example, damaging mutations and copy number variants (microdeletions and microduplications) may affect reproductive system development³³⁶⁻³⁴⁰ and

function³⁴¹⁻³⁴³ as well as fetal, childhood, adolescent and/or adult development and/or function of other organ systems in the body. Indeed, GeneCards³⁴⁴ lists >3,600 gene defects associated with human male infertility and another 3,200+ genes associated with genitourinary birth defects causing abnormal male reproductive development and function. This knowledge will improve clinical diagnosis and treatment.

The potential impact of these genetic findings is in the area of genetic and genomic-based spermiogenesis defects causing teratozoospermia and/or asthenozoospermia (multiple abnormalities of the sperm flagella and primary ciliary dyskinesia). Today, this knowledge is used clinically to counsel patients about their chances for successful ART.^{345, 346} As many of these “infertility” genes are expressed in select other tissues or even broadly throughout the body, infertility may be the “canary in the coal mine” that portends an increased likelihood of other comorbidities. Given the wide range of types of genes required for fertility,³⁴⁷⁻³⁴⁹ it is not surprising that male infertility is associated with other health conditions, such as mortality, malignancies, immune dysfunction, and other non-reproductive disorders.

The impact of certain lifestyles and behaviors remains relatively unknown. For example, vaping and cannabis use are highly prevalent among young adults, but the precise short- and long-term effects of these agents on reproductive health remain unclear.³⁵⁰⁻³⁵³ While obesity and metabolic syndrome can impair male fertility via numerous pathophysiological mechanisms, the ability to restore reproductive potential through weight loss and enhanced metabolic health remains understudied. The emergence of the agonists of glucagon-like peptide-1 receptors (GLP-1 AR) class of drugs are proving to be highly efficacious in treating obesity and type 2 diabetes; the effect of these therapies on reproductive health remains to be determined.

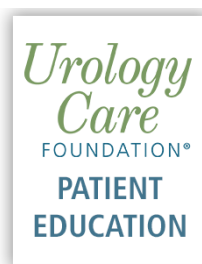
Therapeutic advances for male infertility (except for surgical approaches for obstructive azoospermia and NOA) remain relatively stagnant. However, in the laboratory, novel methods are under development to effectively use spermatogonial stem cells to rejuvenate spermatogenesis after gonadotoxin exposures (such as chemotherapy),³⁵⁴ although potential contamination of spermatogonial stem cells with malignant cells, which must be eliminated before autotransplantation, remain a concern.

Approaches using organ cultures and in vitro systems for spermatogenesis offer additional promise for the treatment of some forms of spermatogenic failure. Qualitative but not quantitative spermatogenesis has been achieved in vitro culminating in live offspring in rodents. With knowledge of the delicate microenvironment needed for completion of spermatogenesis in vitro, researchers are moving closer to achieving this goal, while still maintaining the genetic, genomic, and epigenomic integrity of the sperm.³⁵⁵

Finally, gene therapy approaches targeting the process of spermatogenesis, are advantageous because of the continuous production of sperm throughout the adult lifespan. However, whether germline gene therapy in humans should occur is an ethical question. Questions about whether germline genome editing should be done even for genetic disorders and technical considerations remain problematic.³⁵⁶ Genome editing can result in off-target effects and mosaicism.

In closing, the genomic revolution has placed us at the forefront of vastly improving our diagnostic abilities to define precise etiologies, co-morbidities, and eventually (perhaps) develop medically-based treatments for infertile males to improve not only their fertility potential, but also their overall health. Translation of the newer advances discussed above will be slower but will eventually move from the laboratory to the clinical arena to provide more therapeutic options for males. The future looks promising for improving the health and fertility of the infertile male through precision medicine and the application of advanced technologies.

Tools & Resources



[Male Infertility Web Article](#)

APPENDICES

Appendix I: Male Reproductive Health Physical Examination

The goal of the physical examination is to identify potential etiologies of reproductive impairments, health ailments, or factors that can be optimized to improve health or reproductive success.

General	• Body habitus as overweight obesity is associated with impaired spermatogenesis. • Virilization to assess pubertal development/androgen status • Gynecomastia may be a marker for endocrine disorders
Abdominal exam	• Examination of any scars from prior surgical procedures that may involve the pelvis or impact the urogenital system.
Phallus	• Meatal location as hypospadias/epispadias may make semen deposition in the vagina challenging • Penile plaque as Peyronie's disease may make vaginal intercourse difficult • Penile lesions/ulcers/discharge may be a sign of sexually transmitted infection
Scrotum/Testes	• Examination for prior scars suggesting prior scrotal surgery/trauma • Location as scrotal position of the testes is important for normal function • Size/consistency/contours as a majority of the testis is devoted to spermatogenesis. The exam may also reveal masses consistent with a testicular cancer
Epididymides	• Shape/consistency as normal development should be identified to determine atresia that could be identified by the presence of a <i>CFTR</i> mutation. Induration/dilation could suggest obstruction. Epididymal cysts or spermatoceles may also lead to obstruction.
Vas Deferens	• Shape/consistency as normal development and contour should be confirmed to rule out agenesis as may be seen in the presence of a <i>CFTR</i> mutation or aberrant Wolffian duct embryogenesis • The presence/location of any vasectomy defect or granuloma should also be assessed
Digital Rectal Examination	• Midline prostatic cysts or dilated seminal vesicles may assist in the diagnosis of EDO

ABBREVIATIONS

ABVD	Adriamycin, Bleomycin, Vinblastine, and Dacarbazine
ACOG	American College of Obstetricians and Gynecologists
AMA	American Medical Association
ASCO	American Society of Clinical Oncology
ASRM	American Society for Reproductive Medicine

AUA	American Urological Association
AUAER	American Urological Association Education and Research, Inc.
ASA	Antisperm Antibody
Als	Aromatase Inhibitors
ART	Assisted Reproductive Technology
AZF	Azoospermia Factor
BOD	Board of Directors
BPA	Bisphenol A
CVD	Cardiovascular Disease
CCI	Charlson Comorbidity Index
CBAVD	Congenital Bilateral Absence of the Vas Deferens
CF	Cystic Fibrosis
CFTR	Cystic Fibrosis Transmembrane Conductance Regulator
DNA	Deoxyribonucleic acid
DFI	DNA Fragmentation Index
DEHP	Di-2-ethylhexyl phthalate
EDO	Ejaculatory Duct Obstruction
ECRI	Emergency Care Research Institute
FOE	Failure of Emission
FSH	Follicle-Stimulating Hormone
hCG	Human Chorionic Gonadotropin
HH	Hypogonadotropic Hypogonadism
IB	Immunobead
IVF	In Vitro Fertilization
ICSI	Intracytoplasmic Sperm Injection
IUI	Intrauterine Insemination
LRL	Lower Reference Limits
LH	Luteinizing Hormone
micro-TESE	Microdissection-Testicular Sperm Extraction
MRI	Magnetic Resonance Imaging
NOA	Non-Obstructive Azoospermia
OR	Odds Ratio

PGC	Practice Guidelines Committee
RCTs	Randomized Controlled Trials
RPL	Recurrent Pregnancy Loss
RR	Relative Risk
RE	Retrograde Ejaculation
RPLND	Retroperitoneal Lymph Node Dissection
ROB	Risk of Bias
SQC	Science and Quality Council
SERMs	Selective Estrogen Receptor Modulators
SA	Semen Analysis
SRR	Sperm Retrieval Rates
TL	Telomere
TESE	Testicular Sperm Extraction
TRUS	Transrectal Ultrasonography
TURED	Transurethral Resection of Ejaculatory Ducts
WHO	World Health Organization

REFERENCES

1. Thonneau P, Marchand S, Tallec A et al: Incidence and main causes of infertility in a resident population (1,850,000) of three french regions (1988-1989). *Hum Reprod* 1991; **6**: 811
2. Barratt CLR, Björndahl L, De Jonge CJ et al: The diagnosis of male infertility: An analysis of the evidence to support the development of global who guidance-challenges and future research opportunities. *Hum Reprod Update* 2017; **23**: 660
3. Sigman M, Lipshultz LI and SS H: Office evaluation of the subfertile male, 4 ed. New York: Cambridge University Press, p. 176, 2009
4. Honig SC, Lipshultz LI and Jarow J: Significant medical pathology uncovered by a comprehensive male infertility evaluation. *Fertil Steril* 1994; **62**: 1028
5. Organization WH: Who laboratory manual for the examination and processing of human semen, 5th ed. Geneva, Switzerland: WHO Press, p. 287, 2010
6. Definitions of infertility and recurrent pregnancy loss: A committee opinion. *Fertil Steril* 2020; **113**: 533
7. Improving the reporting of clinical trials of infertility treatments (imprint): Modifying the consort statement. *Fertil Steril* 2014; **102**: 952
8. Chung E, Arafa M, Boitrelle F et al: The new 6th edition of the who laboratory manual for the examination and processing of human semen: Is it a step toward better standard operating procedure? *Asian J Androl* 2022; **24**: 123
9. Boitrelle F, Shah R, Saleh R et al: The sixth edition of the who manual for human semen analysis: A critical review and swot analysis. *Life (Basel)* 2021; **11**
10. Infertility workup for the women's health specialist: Acog committee opinion summary, number 781. *Obstet Gynecol*. 2019; **133**: 1294

11. Optimizing natural fertility: A committee opinion. *Fertil Steril* 2017; **107**: 52
12. Definitions of infertility and recurrent pregnancy loss: A committee opinion. *Fertil Steril* 2013; **99**: 63
13. Jeve YB and Davies W: Evidence-based management of recurrent miscarriages. *J Hum Reprod Sci* 2014; **7**: 159
14. Rowe T: Fertility and a woman's age. *J Reprod Med* 2006; **51**: 157
15. Schwartz D and Mayaux MJ: Female fecundity as a function of age: Results of artificial insemination in 2193 nulliparous women with azoospermic husbands. *Federation cecos. N Engl J Med* 1982; **306**: 404
16. Kumar N and Singh AK: Trends of male factor infertility, an important cause of infertility: A review of literature. *J Hum Reprod Sci* 2015; **8**: 191
17. Eimers JM, te Velde ER, Gerritse R et al: The prediction of the chance to conceive in subfertile couples. *Fertil Steril* 1994; **61**: 44
18. Ramasamy R, Scovell JM, Kovac JR et al: Fluorescence in situ hybridization detects increased sperm aneuploidy in men with recurrent pregnancy loss. *Fertil Steril* 2015; **103**: 906
19. World Health O: Who laboratory manual for the examination and processing of human semen, 5th ed ed. Geneva: World Health Organization, 2010
20. Higgins J: Assessing quality of included studies in cochrane reviews. *The Cochrane Collaboration Methods Groups Newsletter* 2007; **11**
21. Whiting PF, Rutjes AW, Westwood ME et al: Quadas-2: A revised tool for the quality assessment of diagnostic accuracy studies. *Ann Intern Med* 2011; **155**: 529
22. Faraday M, Hubbard H, Kosiak B and Dmochowski R: Staying at the cutting edge: A review and analysis of evidence reporting and grading; the recommendations of the american urological association. *BJU Int* 2009; **104**: 294
23. Fertility problems: Assessment and treatment: National Institute for Health and Care Excellence (UK), p. 142, 2017
24. Infertility workup for the women's health specialist: Acog committee opinion, number 781. *Obstet Gynecol* 2019; **133**: e377
25. Optimizing natural fertility: A committee opinion. *F&S* 2017; **107**: 52
26. Spandorfer SD, Chung PH, Kligman I et al: An analysis of the effect of age on implantation rates. *J Assist Reprod Genet* 2000; **17**: 303
27. Dunson DB, Baird DD and Colombo B: Increased infertility with age in men and women. *Obstet Gynecol* 2004; **103**: 51
28. Guzick DS, Overstreet JW, Factor-Litvak P et al: Sperm morphology, motility, and concentration in fertile and infertile men. *N Engl J Med* 2001; **345**: 1388
29. The optimal evaluation of the infertile male: Aua best practice statement, 2010. Practice committee of the american society for reproductive medicine. Diagnostic evaluation of the infertile male: A committee opinion. *Fertil Steril* 2012; **98**: 294
30. Organization WH: Who laboratory manual for the examination and processing of human semen, 6th ed. Geneva, Switzerland: WHO, p. 276, 2021
31. Campbell MJ, Lotti F, Baldi E et al: Distribution of semen examination results 2020 - a follow up of data collated for the who semen analysis manual 2010. *Andrology* 2021; **9**: 817
32. Cooper TG, Noonan E, von Eckardstein S et al: World health organization reference values for human semen characteristics. *Hum Reprod Update* 2010; **16**: 231
33. Gonzalez D, Narasimman M, Best JC et al: Clinical update on home testing for male fertility. *World J Mens Health* 2021; **39**: 615
34. Cayan S, Erdemir F, Ozbey I et al: Can varicocele surgery significantly change the way couples use assisted reproductive technologies? *J Urol* 2002; **167**: 1749

35. Meng MV, Greene KL and Turek PJ: Surgery or assisted reproduction? A decision analysis of treatment costs in male infertility. *J Urol* 2005; **174**: 1926
36. Schlegel PN: Is assisted reproduction the optimal treatment for varicocele-associated male infertility? A cost-effectiveness analysis. *Urology* 1997; **49**: 83
37. Lira Neto FT, Roque M and Esteves SC: Effect of varicolectomy on sperm deoxyribonucleic acid fragmentation rates in infertile men with clinical varicocele: A systematic review and meta-analysis. *Fertil Steril* 2021; **116**: 696
38. Lee R, Li PS, Goldstein M et al: A decision analysis of treatments for obstructive azoospermia. *Hum Reprod* 2008; **23**: 2043
39. Pavlovich CP and Schlegel PN: Fertility options after vasectomy: A cost-effectiveness analysis. *Fertil Steril* 1997; **67**: 133
40. Kallen B, Finnstrom O, Lindam A et al: Cancer risk in children and young adults conceived by in vitro fertilization. *Pediatrics* 2010; **126**: 270
41. Jensen TK, Jorgensen N, Asklund C et al: Fertility treatment and reproductive health of male offspring: A study of 1,925 young men from the general population. *Am J Epidemiol* 2007; **165**: 583
42. Spector LG, Brown MB, Wantman E et al: Association of in vitro fertilization with childhood cancer in the united states. *JAMA Pediatr* 2019; **173**: e190392
43. Kolettis PN and Sabanegh ES: Significant medical pathology discovered during a male infertility evaluation. *J Urol* 2001; **166**: 178
44. Ventimiglia E, Capogrosso P, Boeri L et al: Infertility as a proxy of general male health: Results of a cross-sectional survey. *Fertil Steril* 2015; **104**: 48
45. Eisenberg ML, Li S, Behr B et al: Relationship between semen production and medical comorbidity. *Fertil Steril* 2015; **103**: 66
46. Bach PV, Patel N, Najari BB et al: Changes in practice patterns in male infertility cases in the united states: The trend toward subspecialization. *Fertil Steril* 2018; **110**: 76
47. Li S, Zheng PS, Ma HM et al: Systematic review of subsequent pregnancy outcomes in couples with parental abnormal chromosomal karyotypes and recurrent pregnancy loss. *Fertil Steril* 2022; **118**: 906
48. Hanif MI, Khan A, Arif A and Shoeb E: Cytogenetic investigation of couples with recurrent spontaneous miscarriages. *Pak J Med Sci* 2019; **35**: 1422
49. Chua SC, Yovich SJ, Hinchliffe PM and Yovich JL: The sperm DNA fragmentation assay with sdf level less than 15% provides a useful prediction for clinical pregnancy and live birth for women aged under 40 years. *J Pers Med* 2023; **13**
50. Lourenço ML, Moura GA, Rocha YM et al: Impact of sperm DNA fragmentation on the clinical outcome of assisted reproduction techniques: A systematic review of the last five years. *JBRA Assist Reprod* 2023; **27**: 282
51. Okubo T, Onda N, Hayashi T et al: Performing a sperm DNA fragmentation test in addition to semen examination based on the who criteria can be a more accurate diagnosis of ivf outcomes. *BMC Urol* 2023; **23**: 78
52. Henkel R, Morris A, Vogiatzi P et al: Predictive value of seminal oxidation-reduction potential analysis for reproductive outcomes of icsi. *Reprod Biomed Online* 2022; **45**: 1007
53. Tang L, Rao M, Yang W et al: Predictive value of the sperm DNA fragmentation index for low or failed ivf fertilization in men with mild-to-moderate asthenozoospermia. *J Gynecol Obstet Hum Reprod* 2021; **50**: 101868
54. Nicopoullos J, Vicens-Morton A, Lewis SEM et al: Novel use of comet parameters of sperm DNA damage may increase its utility to diagnose male infertility and predict live births following both ivf and icsi. *Hum Reprod* 2019; **34**: 1915
55. Fakhrabadi M, Kalantar S, Montazeri F et al: Fish-based sperm aneuploidy screening in male partner of women with a history of recurrent pregnancy loss. *Middle East Fertility Society Journal* 2020; **25**
56. Salonia A, Matloob R, Gallina A et al: Are infertile men less healthy than fertile men? Results of a prospective case control survey. *Eur Urol* 2009; **56**: 1025

57. Oliva A and Multigner L: Chronic epididymitis and grade iii varicocele and their associations with semen characteristics in men consulting for couple infertility. *Asian J Androl* 2018; **20**: 360
58. Cazzaniga W, Capogrosso P, Ventimiglia E et al: High blood pressure is a highly prevalent but unrecognised condition in primary infertile men: Results of a cross-sectional study. *Eur Urol Focus* 2020; **6**: 178
59. Negri L, Benaglia R, Fiamengo B et al: Cancer risk in male factor-infertility. *Placenta* 2008; **29 Suppl B**: 178
60. Hanson HA, Anderson RE, Aston KI et al: Subfertility increases risk of testicular cancer: Evidence from population-based semen samples. *Fertil Steril* 2016; **105**: 322
61. Mancini M, Carmignani L, Gazzano G et al: High prevalence of testicular cancer in azoospermic men without spermatogenesis. *Hum Reprod* 2007; **22**: 1042
62. Raman JD, Nobert CF and Goldstein M: Increased incidence of testicular cancer in men presenting with infertility and abnormal semen analysis. *J Urol* 2005; **174**: 1819
63. Eisenberg ML, Betts P, Herder D et al: Increased risk of cancer among azoospermic men. *Fertil Steril* 2013; **100**: 681
64. Glazer CH, Tøttenborg SS, Giwercman A et al: Male factor infertility and risk of multiple sclerosis: A register-based cohort study. *Mult Scler* 2018; **24**: 1835
65. Glazer CH, Bonde JP, Giwercman A et al: Risk of diabetes according to male factor infertility: A register-based cohort study. *Hum Reprod* 2017; **32**: 1474
66. Bezold G, Politch JA, Kiviat NB et al: Prevalence of sexually transmissible pathogens in semen from asymptomatic male infertility patients with and without leukocytospermia. *Fertil Steril* 2007; **87**: 1087
67. Poppe K, Glinier D, Tournaye H et al: Is systematic screening for thyroid disorders indicated in subfertile men? *Eur J Endocrinol* 2006; **154**: 363
68. Glazer CH, Bonde JP, Eisenberg ML et al: Male infertility and risk of nonmalignant chronic diseases: A systematic review of the epidemiological evidence. *Semin Reprod Med* 2017; **35**: 282
69. Al-Jebari Y, Elenkov A, Wirestrand E et al: Risk of prostate cancer for men fathering through assisted reproduction: Nationwide population based register study. *Bmj* 2019; **366**: l5214
70. Wang NN, Dallas K, Li S et al: The association between varicoceles and vascular disease: An analysis of u.S. Claims data. *Andrology* 2018; **6**: 99
71. Treadwell JR and Oristaglio J: Aua guideline on male infertility evidence report. Edited by ECRI. Linthicum, MD, 2019
72. Bojesen A, Stochholm K, Juul S and Gravholt CH: Socioeconomic trajectories affect mortality in klinefelter syndrome. *J Clin Endocrinol Metab* 2011; **96**: 2098
73. Ishikawa T, Yamaguchi K, Kondo Y et al: Metabolic syndrome in men with klinefelter's syndrome. *Urology* 2008; **71**: 1109
74. Pawlaczyk-Kamieńska T, Borysewicz-Lewicka M, Śniatała R et al: Dental and periodontal manifestations in patients with cystic fibrosis - a systematic review. *J Cyst Fibros* 2019; **18**: 762
75. Chariatte V, Ramseyer P and Cachat F: Uroradiological screening for upper and lower urinary tract anomalies in patients with hypospadias: A systematic literature review. *Evid Based Med* 2013; **18**: 11
76. Akre O, Pettersson A and Richiardi L: Risk of contralateral testicular cancer among men with unilaterally undescended testis: A meta analysis. *Int J Cancer* 2009; **124**: 687
77. Zarotsky V, Huang MY, Carman W et al: Systematic literature review of the risk factors, comorbidities, and consequences of hypogonadism in men. *Andrology* 2014; **2**: 819
78. Kellesarian SV, Malmstrom H, Abduljabbar T et al: "Low testosterone levels in body fluids are associated with chronic periodontitis". *Am J Mens Health* 2017; **11**: 443
79. Radhakrishnan K, Toprac P, O'Hair M et al: Interactive digital e-health game for heart failure self-management: A feasibility study. *Games Health J* 2016; **5**: 366

80. Johnson SL, Dunleavy J, Gemmell NJ and Nakagawa S: Consistent age-dependent declines in human semen quality: A systematic review and meta-analysis. *Ageing Res Rev* 2015; **19**: 22
81. Sartorius GA and Nieschlag E: Paternal age and reproduction. *Hum Reprod Update* 2010; **16**: 65
82. Kong A, Frigge ML, Masson G et al: Rate of de novo mutations and the importance of father's age to disease risk. *Nature* 2012; **488**: 471
83. Jónsson H, Sulem P, Kehr B et al: Parental influence on human germline de novo mutations in 1,548 trios from iceland. *Nature* 2017; **549**: 519
84. Oldereid NB, Wennerholm UB, Pinborg A et al: The effect of paternal factors on perinatal and paediatric outcomes: A systematic review and meta-analysis. *Hum Reprod Update* 2018; **24**: 320
85. du Fossé NA, van der Hoorn MP, van Lith JMM et al: Advanced paternal age is associated with an increased risk of spontaneous miscarriage: A systematic review and meta-analysis. *Hum Reprod Update* 2020; **26**: 650
86. Welcome to reprotox, 2020
87. Bonde JP, Flachs EM, Rimborg S et al: The epidemiologic evidence linking prenatal and postnatal exposure to endocrine disrupting chemicals with male reproductive disorders: A systematic review and meta-analysis. *Hum Reprod Update* 2016; **23**: 104
88. Skakkebaek NE, Rajpert-De Meyts E and Main KM: Testicular dysgenesis syndrome: An increasingly common developmental disorder with environmental aspects. *Hum Reprod* 2001; **16**: 972
89. Mendiola J, Jørgensen N, Andersson AM et al: Are environmental levels of bisphenol a associated with reproductive function in fertile men? *Environ Health Perspect* 2010; **118**: 1286
90. Golub: Metals, fertility and reproductive toxicity. New York, NY, 2006
91. CDC: Lead in drinking water, vol. 2020, 2020
92. Koh DH, Locke SJ, Chen YC et al: Lead exposure in us worksites: A literature review and development of an occupational lead exposure database from the published literature. *Am J Ind Med* 2015; **58**: 605
93. Barbosa F, Jr., Tanus-Santos JE, Gerlach RF and Parsons PJ: A critical review of biomarkers used for monitoring human exposure to lead: Advantages, limitations, and future needs. *Environ Health Perspect* 2005; **113**: 1669
94. Zhang Y, Li S and Li S: Relationship between cadmium content in semen and male infertility: A meta-analysis. *Environ Sci Pollut Res Int* 2019; **26**: 1947
95. Whorton D, Krauss RM, Marshall S and Milby TH: Infertility in male pesticide workers. *Lancet* 1977; **2**: 1259
96. Martenies SE and Perry MJ: Environmental and occupational pesticide exposure and human sperm parameters: A systematic review. *Toxicology* 2013; **307**: 66
97. Zota AR, Calafat AM and Woodruff TJ: Temporal trends in phthalate exposures: Findings from the national health and nutrition examination survey, 2001-2010. *Environ Health Perspect* 2014; **122**: 235
98. Ha yBB, Lenters V, Giwercman A et al: Impact of di-2-ethylhexyl phthalate metabolites on male reproductive function: A systematic review of human evidence. *Curr Environ Health Rep* 2018; **5**: 20
99. Diagnostic evaluation of the infertile male: A committee opinion. *Fertil Steril* 2015; **103**: e18
100. Sigman M and Jarow JP: Endocrine evaluation of infertile men. *Urology* 1997; **50**: 659
101. Ventimiglia E, Capogrosso P, Boeri L et al: Validation of the american society for reproductive medicine guidelines/recommendations in white european men presenting for couple's infertility. *Fertil Steril* 2016; **106**: 1076
102. Olesen IA, Andersson AM, Aksglaede L et al: Clinical, genetic, biochemical, and testicular biopsy findings among 1,213 men evaluated for infertility. *Fertil Steril* 2017; **107**: 74
103. Mulhall JP, Trost LW, Brannigan RE et al: Evaluation and management of testosterone deficiency: Aua guideline. *J Urol* 2018; **200**: 423

104. Schoor RA, Elhanbly S, Niederberger CS and Ross LS: The role of testicular biopsy in the modern management of male infertility. *J Urol* 2002; **167**: 197
105. Corona G, Wu FC, Rastrelli G et al: Low prolactin is associated with sexual dysfunction and psychological or metabolic disturbances in middle-aged and elderly men: The european male aging study (emas). *J Sex Med* 2014; **11**: 240
106. Kamischke A and Nieschlag E: Treatment of retrograde ejaculation and anejaculation. *Hum Reprod Update* 1999; **5**: 448
107. Oates R: Evaluation of the azoospermic male. *Asian J Androl* 2012; **14**: 82
108. Behre HM, Bergmann M, Simoni M and Tuttelmann F: Primary testicular failure. [updated 2015 aug 30]. South Dartmouth, MA: MDText.com, Inc., 2000.
109. Zhao WW, Wu M, Chen F et al: Robertsonian translocations: An overview of 872 robertsonian translocations identified in a diagnostic laboratory in china. *PLoS One* 2015; **10**: e0122647
110. Morel F, Douet-Guilbert N, Le Bris MJ et al: Meiotic segregation of translocations during male gametogenesis. *Int J Androl* 2004; **27**: 200
111. Aksglaede L, Jørgensen N, Skakkebaek NE and Juul A: Low semen volume in 47 adolescents and adults with 47, xxy klinefelter or 46, xx male syndrome. *Int J Androl* 2009; **32**: 376
112. Laron Z, Dickerman Z, Zamir R and Galatzer A: Paternity in klinefelter's syndrome--a case report. *Arch Androl* 1982; **8**: 149
113. Terzoli G, Lalatta F, Lobbiani A et al: Fertility in a 47, xxy patient: Assessment of biological paternity by deoxyribonucleic acid fingerprinting. *Fertil Steril* 1992; **58**: 821
114. Lin YM, Huang WJ, Lin JS and Kuo PL: Progressive depletion of germ cells in a man with nonmosaic klinefelter's syndrome: Optimal time for sperm recovery. *Urology* 2004; **63**: 380
115. Ichioka K, Utsunomiya N, Kohei N et al: Adult onset of declining spermatogenesis in a man with nonmosaic klinefelter's syndrome. *Fertil Steril* 2006; **85**: 1511.e1
116. Sabbaghian M, Mohseni Meybodi A, Rafeae A et al: Sperm retrieval rate and reproductive outcome of infertile men with azoospermia factor c deletion. *Andrologia* 2018; **50**: e13052
117. Yuen W, Golin AP, Flannigan R and Schlegel PN: Histology and sperm retrieval among men with y chromosome microdeletions. *Transl Androl Urol* 2021; **10**: 1442
118. Rabinowitz MJ, Huffman PJ, Haney NM and Kohn TP: Y-chromosome microdeletions: A review of prevalence, screening, and clinical considerations. *Appl Clin Genet* 2021; **14**: 51
119. Minhas S, Bettocchi C, Boeri L et al: European association of urology guidelines on male sexual and reproductive health: 2021 update on male infertility. *Eur Urol* 2021; **80**: 603
120. Kohn TP, Kohn JR, Owen RC and Coward RM: The prevalence of y-chromosome microdeletions in oligozoospermic men: A systematic review and meta-analysis of european and north american studies. *Eur Urol* 2019; **76**: 626
121. Tang D, Liu W, Li G et al: Normal fertility with deletion of sy84 and sy86 in azfa region. *Andrology* 2020; **8**: 332
122. Alksere B, Berzina D, Dudorova A et al: Case of inherited partial azfa deletion without impact on male fertility. *Case Rep Genet* 2019; **2019**: 3802613
123. Stouffs K, Vloeberghs V, Gheldof A et al: Are azfb deletions always incompatible with sperm production? *Andrology* 2017; **5**: 691
124. Hopps CV, Mielnik A, Goldstein M et al: Detection of sperm in men with y chromosome microdeletions of the azfa, azfb and azfc regions. *Hum Reprod* 2003; **18**: 1660
125. Krausz C, Hoefsloot L, Simoni M and Tüttelmann F: Eaa/emqn best practice guidelines for molecular diagnosis of chromosomal microdeletions: State-of-the-art 2013. *Andrology* 2014; **2**: 5

126. Reddy MM and Stutts MJ: Status of fluid and electrolyte absorption in cystic fibrosis. *Cold Spring Harb Perspect Med* 2013; **3**: a009555
127. Gaillard DA, Carre-Pigeon F and Lallemant A: Normal vas deferens in fetuses with cystic fibrosis. *J Urol* 1997; **158**: 1549
128. Mak V, Zielenski J, Tsui LC et al: Proportion of cystic fibrosis gene mutations not detected by routine testing in men with obstructive azoospermia. *Jama* 1999; **281**: 2217
129. Chillon M, Casals T, Mercier B et al: Mutations in the cystic fibrosis gene in patients with congenital absence of the vas deferens. *N Engl J Med* 1995; **332**: 1475
130. Yu J, Chen Z, Ni Y and Li Z: Cftr mutations in men with congenital bilateral absence of the vas deferens (cbavd): A systemic review and meta-analysis. *Hum Reprod* 2012; **27**: 25
131. Mehdizadeh Hakkak A, Keramatipour M, Talebi S et al: Analysis of cftr gene mutations in children with cystic fibrosis, first report from north-east of iran. *Iran J Basic Med Sci* 2013; **16**: 917
132. Alper OM, Wong LJ, Young S et al: Identification of novel and rare mutations in california hispanic and african american cystic fibrosis patients. *Hum Mutat* 2004; **24**: 353
133. Bobadilla JL, Macek M, Jr., Fine JP and Farrell PM: Cystic fibrosis: A worldwide analysis of cftr mutations--correlation with incidence data and application to screening. *Hum Mutat* 2002; **19**: 575
134. Palomaki GE, FitzSimmons SC and Haddow JE: Clinical sensitivity of prenatal screening for cystic fibrosis via cftr carrier testing in a united states panethnic population. *Genet Med* 2004; **6**: 405
135. Schrijver I, Pique L, Graham S et al: The spectrum of cftr variants in nonwhite cystic fibrosis patients: Implications for molecular diagnostic testing. *J Mol Diagn* 2016; **18**: 39
136. Patat O, Pagin A, Siegfried A et al: Truncating mutations in the adhesion g protein-coupled receptor g2 gene *adgrg2* cause an x-linked congenital bilateral absence of vas deferens. *Am J Hum Genet* 2016; **99**: 437
137. Acog committee opinion no. 762: Prepregnancy counseling. *Obstet Gynecol* 2019; **133**: e78
138. Bradley CK, McArthur SJ, Gee AJ et al: Intervention improves assisted conception intracytoplasmic sperm injection outcomes for patients with high levels of sperm DNA fragmentation: A retrospective analysis. *Andrology* 2016; **4**: 903
139. Deng N, Haney NM, Kohn TP et al: The effect of shift work on urogenital disease: A systematic review. *Curr Urol Rep* 2018; **19**: 57
140. Mohamed EE and Mohamed MA: Effect of sperm chromatin condensation on the outcome of intrauterine insemination in patients with male factor infertility. *J Reprod Med* 2012; **57**: 421
141. Simon L, Zini A, Dyachenko A et al: A systematic review and meta-analysis to determine the effect of sperm DNA damage on in vitro fertilization and intracytoplasmic sperm injection outcome. *Asian J Androl* 2017; **19**: 80
142. Dong J, Lv Y, Zhu G et al: Effect of sperm DNA fragmentation on the clinical outcomes of two assisted reproduction methods: Ivf and icsi. *Int J Clin Exp Med* 2017; **10**: 11812
143. Esteves SC, Roque M, Bradley CK and Garrido N: Reproductive outcomes of testicular versus ejaculated sperm for intracytoplasmic sperm injection among men with high levels of DNA fragmentation in semen: Systematic review and meta-analysis. *Fertil Steril* 2017; **108**: 456
144. Ayad BM, Horst GV and Plessis SSD: Revisiting the relationship between the ejaculatory abstinence period and semen characteristics. *Int J Fertil Steril* 2018; **11**: 238
145. Heidenreich A, Bonfig R, Wilbert DM et al: Risk factors for antisperm antibodies in infertile men. *Am J Reprod Immunol* 1994; **31**: 69
146. Munuce MJ, Berta CL, Pauluzzi F and Caille AM: Relationship between antisperm antibodies, sperm movement, and semen quality. *Urol Int* 2000; **65**: 200
147. Lee R, Goldstein M, Ullery BW et al: Value of serum antisperm antibodies in diagnosing obstructive azoospermia. *J Urol* 2009; **181**: 264

148. Bollendorf A, Check JH, Katsoff D and Fedele A: The use of chymotrypsin/galactose to treat spermatozoa bound with anti-sperm antibodies prior to intra-uterine insemination. *Hum Reprod* 1994; **9**: 484
149. Check JH, Hourani W, Check ML et al: Effect of treating antibody-coated sperm with chymotrypsin on pregnancy rates following iui as compared to outcome of ivf/icsi. *Arch Androl* 2004; **50**: 93
150. Gekas J, Thepot F, Turleau C et al: Chromosomal factors of infertility in candidate couples for icsi: An equal risk of constitutional aberrations in women and men. *Hum Reprod* 2001; **16**: 82
151. Check JH, Graziano V, Cohen R et al: Effect of an abnormal sperm chromatin structural assay (scca) on pregnancy outcome following (ivf) with icsi in previous ivf failures. *Arch Androl* 2005; **51**: 121
152. McQueen DB, Zhang J and Robins JC: Sperm DNA fragmentation and recurrent pregnancy loss: A systematic review and meta-analysis. *Fertil Steril* 2019; **112**: 54
153. Kamkar N, Ramezanali F and Sabbaghian M: The relationship between sperm DNA fragmentation, free radicals and antioxidant capacity with idiopathic repeated pregnancy loss. *Reprod Biol* 2018; **18**: 330
154. Carlini T, Paoli D, Pelloni M et al: Sperm DNA fragmentation in italian couples with recurrent pregnancy loss. *Reprod Biomed Online* 2017; **34**: 58
155. Talebi AR, Vahidi S, Aflatoonian A et al: Cytochemical evaluation of sperm chromatin and DNA integrity in couples with unexplained recurrent spontaneous abortions. *Andrologia* 2012; **44 Suppl 1**: 462
156. Egozcue S, Blanco J, Vendrell JM et al: Human male infertility: Chromosome anomalies, meiotic disorders, abnormal spermatozoa and recurrent abortion. *Hum Reprod Update* 2000; **6**: 93
157. Rubio C, Simón C, Blanco J et al: Implications of sperm chromosome abnormalities in recurrent miscarriage. *J Assist Reprod Genet* 1999; **16**: 253
158. Harton GL and Tempest HG: Chromosomal disorders and male infertility. *Asian J Androl* 2012; **14**: 32
159. Hassold T and Hunt P: To err (meiotically) is human: The genesis of human aneuploidy. *Nat Rev Genet* 2001; **2**: 280
160. Kohn TP, Kohn JR, Darilek S et al: Genetic counseling for men with recurrent pregnancy loss or recurrent implantation failure due to abnormal sperm chromosomal aneuploidy. *J Assist Reprod Genet* 2016; **33**: 571
161. Rodrigo L, Rubio C, Peinado V et al: Testicular sperm from patients with obstructive and nonobstructive azoospermia: Aneuploidy risk and reproductive prognosis using testicular sperm from fertile donors as control samples. *Fertil Steril* 2011; **95**: 1005
162. Jarow JP, Ogle SR and Eskew LA: Seminal improvement following repair of ultrasound detected subclinical varicoceles. *J Urol* 1996; **155**: 1287
163. Infertility in the male, 4 ed. Cambridge: Cambridge University Press, 2009
164. Lotti F and Maggi M: Ultrasound of the male genital tract in relation to male reproductive health. *Hum Reprod Update* 2015; **21**: 56
165. Singh R, Hamada AJ, Bukavina L and Agarwal A: Physical deformities relevant to male infertility. *Nat Rev Urol* 2012; **9**: 156
166. Avellino GJ, Lipshultz LI, Sigman M and Hwang K: Transurethral resection of the ejaculatory ducts: Etiology of obstruction and surgical treatment options. *Fertil Steril* 2019; **111**: 427
167. Biyani CS, Cartledge J and Janetschek G: Varicocele. *BMJ Clin Evid* 2009; ~
168. Bate J: Symptomatic varicocele. *Journal of Urology* 1927; **18**: 649
169. Cheungpasitporn W, Horne JM and Howarth CB: Adrenocortical carcinoma presenting as varicocele and renal vein thrombosis: A case report. *J Med Case Rep* 2011; **5**: 337
170. Spittel JA, Jr., Deweerd JH and Shick RM: Acute varicocele: A vascular clue to renal tumor. *Proc Staff Meet Mayo Cl* 1959; **34**: 134

171. Elmer DeWitt M, Greene DJ, Gill B et al: Isolated right varicocele and incidence of associated cancer. *Urology* 2018; **117**: 82
172. Kolettis PN and Sandlow JI: Clinical and genetic features of patients with congenital unilateral absence of the vas deferens. *Urology* 2002; **60**: 1073
173. Schlegel PN, Shin D and Goldstein M: Urogenital anomalies in men with congenital absence of the vas deferens. *J Urol* 1996; **155**: 1644
174. Weiske WH, Salzler N, Schroeder-Printzen I and Weidner W: Clinical findings in congenital absence of the vasa deferentia. *Andrologia* 2000; **32**: 13
175. Lane VA, Scammell S, West N and Murthi GV: Congenital absence of the vas deferens and unilateral renal agenesis: Implications for patient and family. *Pediatr Surg Int* 2014; **30**: 733
176. Wang J, Xia SJ, Liu ZH et al: Inguinal and subinguinal micro-varicocelectomy, the optimal surgical management of varicocele: A meta-analysis. *Asian J Androl* 2015; **17**: 74
177. Kirby EW, Wiener LE, Rajanahally S et al: Undergoing varicocele repair before assisted reproduction improves pregnancy rate and live birth rate in azoospermic and oligospermic men with a varicocele: A systematic review and meta-analysis. *Fertil Steril* 2016; **106**: 1338
178. Kim HJ, Seo JT, Kim KJ et al: Clinical significance of subclinical varicocelectomy in male infertility: Systematic review and meta-analysis. *Andrologia* 2016; **48**: 654
179. Ron-El R, Strassburger D, Friedler S et al: Extended sperm preparation: An alternative to testicular sperm extraction in non-obstructive azoospermia. *Hum Reprod* 1997; **12**: 1222
180. Schlegel PN and Goldstein M: Alternate indications for varicocele repair: Non-obstructive azoospermia, pain, androgen deficiency and progressive testicular dysfunction. *Fertil Steril* 2011; **96**: 1288
181. Schlegel PN and Kaufmann J: Role of varicocelectomy in men with nonobstructive azoospermia. *Fertil Steril* 2004; **81**: 1585
182. Ramasamy R, Yagan N and Schlegel PN: Structural and functional changes to the testis after conventional versus microdissection testicular sperm extraction. *Urology* 2005; **65**: 1190
183. Bernie AM, Mata DA, Ramasamy R and Schlegel PN: Comparison of microdissection testicular sperm extraction, conventional testicular sperm extraction, and testicular sperm aspiration for nonobstructive azoospermia: A systematic review and meta-analysis. *Fertil Steril* 2015; **104**: ~
184. Yu Z, Wei Z, Yang J et al: Comparison of intracytoplasmic sperm injection outcome with fresh versus frozen-thawed testicular sperm in men with nonobstructive azoospermia: A systematic review and meta-analysis. *J Assist Reprod Genet* 2018; **35**: 1247
185. Nicopoullos JDM, Gilling-Smith C, Almeida PA et al: Use of surgical sperm retrieval in azoospermic men: A meta-analysis. *Fertil Steril* 2004; **82**: 691
186. Marmar JL, Sharlip I and Goldstein M: Results of vasovasostomy or vasoepididymostomy after failed percutaneous epididymal sperm aspirations. *J Urol* 2008; **179**: 1506
187. Chan PT and Libman J: Feasibility of microsurgical reconstruction of the male reproductive tract after percutaneous epididymal sperm aspiration (pesa). *Can J Urol* 2003; **10**: 2070
188. Karavani G, Juvet TSJ, Lau S et al: Improved sperm DNA fragmentation levels in infertile men following very short abstinence of 3-4 hours. *Transl Androl Urol* 2023; **12**: 1487
189. Hervás I, Gil Julia M, Rivera-Egea R et al: Switching to testicular sperm after a previous icsi failure with ejaculated sperm significantly improves blastocyst quality without increasing aneuploidy risk. *J Assist Reprod Genet* 2022; **39**: 2275
190. Zhao G, Jiang X, Zheng Y et al: Outcomes comparison of testicular versus ejaculated sperm for intracytoplasmic sperm injection in infertile men with high DNA fragmentation: Updated systematic review and meta-analysis. *Transl Androl Urol* 2023; **12**: 1785

191. Zhang J, Xue H, Qiu F et al: Testicular spermatozoon is superior to ejaculated spermatozoon for intracytoplasmic sperm injection to achieve pregnancy in infertile males with high sperm DNA damage. *Andrologia* 2019; **51**: e13175
192. Loloi J, Petrella F, Kresch E et al: The effect of sperm DNA fragmentation on male fertility and strategies for improvement: A narrative review. *Urology* 2022; **168**: 3
193. Sigman M: Introduction: Ejaculatory problems and male infertility. *Fertil Steril* 2015; **104**: 1049
194. Valerie U, De BS, De BM et al: Pregnancy after vasectomy: Surgical reversal or assisted reproduction? *Hum Reprod* 2018; **33**: 1218
195. Herrel LA, Goodman M, Goldstein M and Hsiao W: Outcomes of microsurgical vasovasostomy for vasectomy reversal: A meta-analysis and systematic review. *Urology* 2015; **85**: 819
196. Belker AM, Thomas AJ, Jr., Fuchs EF et al: Results of 1,469 microsurgical vasectomy reversals by the vasovasostomy study group. *J Urol* 1991; **145**: 505
197. Engin G: Transrectal us-guided seminal vesicle aspiration in the diagnosis of partial ejaculatory duct obstruction. *Diagn Interv Radiol* 2012; **18**: 488
198. Jarow JP: Transrectal ultrasonography of infertile men. *Fertil Steril* 1993; **60**: 1035
199. Meacham RB, Hellerstein DK and Lipshultz LI: Evaluation and treatment of ejaculatory duct obstruction in the infertile male. *Fertil Steril* 1993; **59**: 393
200. Turek PJ, Magana JO and Lipshultz LI: Semen parameters before and after transurethral surgery for ejaculatory duct obstruction. *J Urol* 1996; **155**: 1291
201. Kadioglu A, Cayan S, Tefekli A et al: Does response to treatment of ejaculatory duct obstruction in infertile men vary with pathology? *Fertil Steril* 2001; **76**: 138
202. Purohit RS, Wu DS, Shinohara K and Turek PJ: A prospective comparison of 3 diagnostic methods to evaluate ejaculatory duct obstruction. *J Urol* 2004; **171**: 232
203. Aggour A, Mostafa H and Maged W: Endoscopic management of ejaculatory duct obstruction. *Int Urol Nephrol* 1998; **30**: 481
204. Tu XA, Zhuang JT, Zhao L et al: Transurethral bipolar plasma kinetic resection of ejaculatory duct for treatment of ejaculatory duct obstruction. *J Xray Sci Technol* 2013; **21**: 293
205. Schroeder-Printzen I, Ludwig M, Kohn F and Weidner W: Surgical therapy in infertile men with ejaculatory duct obstruction: Technique and outcome of a standardized surgical approach. *Hum Reprod* 2000; **15**: 1364
206. El-Assmy A, El-Tholoth H, Abouelkheir RT and Abou-El-Ghar ME: Transurethral resection of ejaculatory duct in infertile men: Outcome and predictors of success. *Int Urol Nephrol* 2012; **44**: 1623
207. Netto NR, Jr., Esteves SC and Neves PA: Transurethral resection of partially obstructed ejaculatory ducts: Seminal parameters and pregnancy outcomes according to the etiology of obstruction. *J Urol* 1998; **159**: 2048
208. Cohen J, Edwards R, Fehilly C et al: In vitro fertilization: A treatment for male infertility. *Fertil Steril* 1985; **43**: 422
209. Sunderam S ZY, Jewett A, Mardovich S, Kissin DM: State-specific assisted reproductive technology surveillance, united states: 2021 data brief. Centers for disease control and prevention, us dept of health and human services. 2023;
210. Finkelstein JS, Whitcomb RW, O'Dea LS et al: Sex steroid control of gonadotropin secretion in the human male. I. Effects of testosterone administration in normal and gonadotropin-releasing hormone-deficient men. *J Clin Endocrinol Metab* 1991; **73**: 609
211. Oliveira LM, Seminara SB, Beranova M et al: The importance of autosomal genes in kallmann syndrome: Genotype-phenotype correlations and neuroendocrine characteristics. *J Clin Endocrinol Metab* 2001; **86**: 1532
212. Gianetti E, Hall JE, Au MG et al: When genetic load does not correlate with phenotypic spectrum: Lessons from the gnrh receptor (gnrhr). *J Clin Endocrinol Metab* 2012; **97**: E1798

213. Pitteloud N, Crowley WF, Jr. and Balasubramanian R: Isolated gonadotropin-releasing hormone deficiency (idiopathic hypogonadotropic hypogonadism). Edited by P. J. Snyder and A. M. Matsumoto: UpToDate, 2020
214. Nachtigall LB, Boepple PA, Pralong FP and Crowley WF, Jr.: Adult-onset idiopathic hypogonadotropic hypogonadism--a treatable form of male infertility. *N Engl J Med* 1997; **336**: 410
215. Rohayem J, Hauffa BP, Zacharin M et al: Testicular growth and spermatogenesis: New goals for pubertal hormone replacement in boys with hypogonadotropic hypogonadism? -a multicentre prospective study of hcg/rfsh treatment outcomes during adolescence. *Clin Endocrinol (Oxf)* 2017; **86**: 75
216. Burris AS, Rodbard HW, Winters SJ and Sherins RJ: Gonadotropin therapy in men with isolated hypogonadotropic hypogonadism: The response to human chorionic gonadotropin is predicted by initial testicular size. *J Clin Endocrinol Metab* 1988; **66**: 1144
217. Miyagawa Y, Tsujimura A, Matsumiya K et al: Outcome of gonadotropin therapy for male hypogonadotropic hypogonadism at university affiliated male infertility centers: A 30-year retrospective study. *J Urol* 2005; **173**: 2072
218. Liu PY, Baker HW, Jayadev V et al: Induction of spermatogenesis and fertility during gonadotropin treatment of gonadotropin-deficient infertile men: Predictors of fertility outcome. *J Clin Endocrinol Metab* 2009; **94**: 801
219. Whitten SJ, Nangia AK and Kolettis PN: Select patients with hypogonadotropic hypogonadism may respond to treatment with clomiphene citrate. *Fertil Steril* 2006; **86**: 1664
220. Chehab M, Madala A and Trussell JC: On-label and off-label drugs used in the treatment of male infertility. *Fertil Steril* 2015; **103**: 595
221. Fraietta R, Zylberstejn DS and Esteves SC: Hypogonadotropic hypogonadism revisited. *Clinics (Sao Paulo)* 2013; **68 Suppl 1**: 81
222. Zumoff B, Miller LK and Strain GW: Reversal of the hypogonadotropic hypogonadism of obese men by administration of the aromatase inhibitor testolactone. *Metabolism* 2003; **52**: 1126
223. de Boer H, Verschoor L, Ruinemans-Koerts J and Jansen M: Letrozole normalizes serum testosterone in severely obese men with hypogonadotropic hypogonadism. *Diabetes Obes Metab* 2005; **7**: 211
224. Contraceptive efficacy of testosterone-induced azoospermia in normal men. World health organization task force on methods for the regulation of male fertility. *Lancet* 1990; **336**: 955
225. Liu PY, Swerdloff RS, Christenson PD et al: Rate, extent, and modifiers of spermatogenic recovery after hormonal male contraception: An integrated analysis. *Lancet* 2006; **367**: 1412
226. Ledesma BR, Weber A, Venigalla G et al: Fertility outcomes in men with prior history of anabolic steroid use. *Fertil Steril* 2023; **120**: 1203
227. Ramasamy R, Armstrong JM and Lipshultz LI: Preserving fertility in the hypogonadal patient: An update. *Asian J Androl* 2015; **17**: 197
228. Ramasamy R, Masterson TA, Best JC et al: Effect of natesto on reproductive hormones, semen parameters and hypogonadal symptoms: A single center, open label, single arm trial. *J Urol* 2020; **204**: 557
229. Kavoussi PK, Machen GL, Gilkey MS et al: Converting men from clomiphene citrate to natesto for hypogonadism improves libido, maintains semen parameters, and reduces estradiol. *Urology* 2021; **148**: 141
230. Bayrak A, Saadat P, Mor E et al: Pituitary imaging is indicated for the evaluation of hyperprolactinemia. *Fertil Steril* 2005; **84**: 181
231. Vilar L, Vilar CF, Lyra R and Freitas MDC: Pitfalls in the diagnostic evaluation of hyperprolactinemia. *Neuroendocrinology* 2019; **109**: 7
232. Famini P, Maya MM and Melmed S: Pituitary magnetic resonance imaging for sellar and parasellar masses: Ten-year experience in 2598 patients. *J Clin Endocrinol Metab* 2011; **96**: 1633
233. Melmed S, Casanueva FF, Hoffman AR et al: Diagnosis and treatment of hyperprolactinemia: An endocrine society clinical practice guideline. *J Clin Endocrinol Metab* 2011; **96**: 273

234. Snyder PJ: Clinical manifestations and evaluation of hyperprolactinemia. Edited by D. S. Cooper: UpToDate, vol. 2020
235. Molitch ME: Diagnosis and treatment of pituitary adenomas: A review. *Jama* 2017; **317**: 516
236. Honegger J, Nasi-Kordhishti I, Aboutaha N and Giese S: Surgery for prolactinomas: A better choice? *Pituitary* 2020; **23**: 45
237. Chua ME, Escusa KG, Luna S et al: Revisiting oestrogen antagonists (clomiphene or tamoxifen) as medical empiric therapy for idiopathic male infertility: A meta-analysis. *Andrology* 2013; **1**: 749
238. Cannarella R, Condorelli RA, Mongioia^a LM et al: Effects of the selective estrogen receptor modulators for the treatment of male infertility: A systematic review and meta-analysis. *Expert Opin Pharmacother* 2019; **20**: 1517
239. Steiner AZ, Hansen KR, Barnhart KT et al: The effect of antioxidants on male factor infertility: The males, antioxidants, and infertility (moxi) randomized clinical trial. *Fertil Steril* 2020; **113**: 552
240. Schisterman EF, Sjaarda LA, Clemons T et al: Effect of folic acid and zinc supplementation in men on semen quality and live birth among couples undergoing infertility treatment: A randomized clinical trial. *Jama* 2020; **323**: 35
241. Santi D, Granata ARM and Simoni M: Fsh treatment of male idiopathic infertility improves pregnancy rate: A meta-analysis. *Endocr Connect* 2015; **4**: R46
242. Attia AM, Al-Inany HG, Farquhar C and Proctor M: Gonadotrophins for idiopathic male factor subfertility. *Cochrane Database Syst Rev* 2007: CD005071
243. Ding YM, Zhang XJ, Li JP et al: Treatment of idiopathic oligozoospermia with recombinant human follicle-stimulating hormone: A prospective, randomized, double-blind, placebo-controlled clinical study in chinese population. *Clin Endocrinol* 2015; **83**: 866
244. Pozzi E, Venigalla G, Raymo A et al: Eligibility for the medical therapy among men with non-obstructive azoospermia-findings from a multi-centric cross-sectional study. *Andrology* 2024: online ahead of print
245. Hussein A, Ozgok Y, Ross L et al: Optimization of spermatogenesis-regulating hormones in patients with non-obstructive azoospermia and its impact on sperm retrieval: A multicentre study. *BJU Int* 2013; **111**: E110
246. Cavallini G, Biagiotti G and Bolzon E: Multivariate analysis to predict letrozole efficacy in improving sperm count of non-obstructive azoospermic and cryptozoospermic patients: A pilot study. *Asian J Androl* 2013; **15**: 806
247. Gül Ü and Turunç T: The effect of human chorionic gonadotropin treatment before testicular sperm extraction in non-obstructive azoospermia. *J Clin Anal Med* 2016; **7**: 55
248. Aydos K, AonIA C, Demirel LC et al: The effect of pure fsh administration in non-obstructive azoospermic men on testicular sperm retrieval. *Eur J Obstet Gynecol Reprod Biol* 2003; **108**: 54
249. Meistrich ML: Effects of chemotherapy and radiotherapy on spermatogenesis in humans. *Fertil Steril* 2013; **100**: 1180
250. Lu CC and Meistrich ML: Cytotoxic effects of chemotherapeutic drugs on mouse testis cells. *Cancer Res* 1979; **39**: 3575
251. Miguel F, Da Cunha MF, Meistrich ML and Mohamed MA: Temporary effects of a msa (4'-(9-acridinylamino) methanesulfon-m-aniside) chemotherapy on spermatogenesis. *Cancer* 1982; **49**: 2459
252. Rowley MJ, Leach DR, Warner GA and Heller CG: Effect of graded doses of ionizing radiation on the human testis. *Radiat Res* 1974; **59**: 665
253. Howell SJ and Shalet SM: Spermatogenesis after cancer treatment: Damage and recovery. *J Natl Cancer Inst Monogr* 2005: 12
254. Hansen PV, Trykker H, Svennekjaer IL and Hvolby J: Long-term recovery of spermatogenesis after radiotherapy in patients with testicular cancer. *Radiother Oncol* 1990; **18**: 117
255. Meistrich M and Beek M: Radiation sensitivity of the human testis, vol. 14, pp. 227-268, 1990
256. Jacob A, Barker H, Goodman A and Holmes J: Recovery of spermatogenesis following bone marrow transplantation. *Bone Marrow Transplant* 1998; **22**: 277

257. Sanders JE, Hawley J, Levy W et al: Pregnancies following high-dose cyclophosphamide with or without high-dose busulfan or total-body irradiation and bone marrow transplantation. *Blood* 1996; **87**: 3045
258. Green DM, Kawashima T, Stovall M et al: Fertility of male survivors of childhood cancer: A report from the childhood cancer survivor study. *J Clin Oncol* 2010; **28**: 332
259. Tomlinson M, Meadows J, Kohut T et al: Review and follow-up of patients using a regional sperm cryopreservation service: Ensuring that resources are targeted to those patients most in need. *Andrology* 2015; **3**: 709
260. Brydoy M, Fosså SD, Klepp O et al: Sperm counts and endocrinological markers of spermatogenesis in long-term survivors of testicular cancer. *Br J Cancer* 2012; **107**: 1833
261. Isaksson S, Eberhard J, Ståhl O et al: Inhibin b concentration is predictive for long-term azoospermia in men treated for testicular cancer. *Andrology* 2014; **2**: 252
262. Gandini L, SgrAý P, Lombardo F et al: Effect of chemo- or radiotherapy on sperm parameters of testicular cancer patients. *Hum Reprod* 2006; **21**: 2882
263. Nurmio M, Keros V, Lahteenmäki P et al: Effect of childhood acute lymphoblastic leukemia therapy on spermatogonia populations and future fertility. *J Clin Endocrinol Metab* 2009; **94**: 2119
264. Stukenborg JB, Alves-Lopes JP, Kurek M et al: Spermatogonial quantity in human prepubertal testicular tissue collected for fertility preservation prior to potentially sterilizing therapy. *Hum Reprod* 2018; **33**: 1677
265. Meistrich ML, Chawla SP, Da Cunha MF et al: Recovery of sperm production after chemotherapy for osteosarcoma. *Cancer* 1989; **63**: 2115
266. Russell LB, Hunsicker PR, Johnson DK and Shelby MD: Unlike other chemicals, etoposide (a topoisomerase-ii inhibitor) produces peak mutagenicity in primary spermatocytes of the mouse. *Mutat Res* 1998; **400**: 279
267. Schultheis B, Nijmeijer BA, Yin H et al: Imatinib mesylate at therapeutic doses has no impact on folliculogenesis or spermatogenesis in a leukaemic mouse model. *Leuk Res* 2012; **36**: 271
268. Brydøy M, Fosså SD, Dahl O and Bjørø T: Gonadal dysfunction and fertility problems in cancer survivors. *Acta Oncol* 2007; **46**: 480
269. Namekawa T, Imamoto T, Kato M et al: Testicular function among testicular cancer survivors treated with cisplatin-based chemotherapy. *Reprod Med Biol* 2016; **15**: 175
270. Bahadur G, Ozturk O, Muneer A et al: Semen quality before and after gonadotoxic treatment. *Hum Reprod* 2005; **20**: 774
271. Spermon JR, Ramos L, Wetzels AMM et al: Sperm integrity pre- and post-chemotherapy in men with testicular germ cell cancer. *Hum Reprod* 2006; **21**: 1781
272. Bohlen D, Burkhard FC, Mills R et al: Fertility and sexual function following orchiectomy and 2 cycles of chemotherapy for stage i high risk nonseminomatous germ cell cancer. *J Urol* 2001; **165**: 441
273. Schrader M, Muller M, Straub B and Miller K: Testicular sperm extraction in azoospermic patients with gonadal germ cell tumors prior to chemotherapy--a new therapy option. *Asian J Androl* 2002; **4**: 9
274. Rafsanjani KA, Faranoush M, Hedayatiasl AA and Vossough P: Gonadal function and fertility in males survivors treated for hodgkin's disease in iran. *Saudi Med J* 2007; **28**: 1690
275. Hobbie WL, Ginsberg JP, Ogle SK et al: Fertility in males treated for hodgkins disease with copp/abv hybrid. *Pediatr Blood Cancer* 2005; **44**: 193
276. Arush MWB, Solt I, Lightman A et al: Male gonadal function in survivors of childhood hodgkin and non-hodgkin lymphoma. *Pediatr Hematol Oncol* 2000; **17**: 239
277. Romerius P, Ståhl O, MoA®ll C et al: High risk of azoospermia in men treated for childhood cancer. *Int J Androl* 2011; **34**: 69
278. Van Beek RD, Smit M, Van Den Heuvel-Eibrink MM et al: Inhibin b is superior to fsh as a serum marker for spermatogenesis in men treated for hodgkin's lymphoma with chemotherapy during childhood. *Hum Reprod* 2007;

279. Paoli D, Rizzo F, Fiore G et al: Spermatogenesis in hodgkin's lymphoma patients: A retrospective study of semen quality before and after different chemotherapy regimens. *Hum Reprod* 2016; **31**: 263
280. Grigg AP, McLachlan R, Zajac J and Szer J: Reproductive status in long-term bone marrow transplant survivors receiving busulfan-cyclophosphamide (120 mg/kg). *Bone Marrow Transplant* 2000; **26**: 1089
281. Green DM, Zhu L, Wang M et al: Effect of cranial irradiation on sperm concentration of adult survivors of childhood acute lymphoblastic leukemia: A report from the st. Jude lifetime cohort study. *Hum Reprod* 2017; **32**: 1192
282. Rendtorff R, Beyer M, Ma-ller A et al: Low inhibin b levels alone are not a reliable marker of dysfunctional spermatogenesis in childhood cancer survivors. *Andrologia* 2012; **44 Suppl 1**: 219
283. Thomson AB, Campbell AJ, Irvine DS et al: Semen quality and spermatozoal DNA integrity in survivors of childhood cancer: A case-control study. *Lancet* 2002; **360**: 361
284. Andreu JAL, FernAndez PJ, FerrA-s IT et al: Persistent altered spermatogenesis in long-term childhood cancer survivors. *Pediatr Hematol Oncol* 2000; **17**: 21
285. Relander T, Cavallin-StA hl E, Garwicz S et al: Gonadal and sexual function in men treated for childhood cancer. *Med Pediatr Oncol* 2000; **35**: 52
286. Lahteenmaki PM, Arola M, Suominen J et al: Male reproductive health after childhood cancer. *Acta Paediatr* 2008; **97**: 935
287. Meistrich ML: Risks of genetic damage in offspring conceived using sperm produced during chemotherapy or radiotherapy. *Andrology* 2019;
288. Russell LB, Hunsicker PR and Russell WL: Comparison of the genetic effects of equimolar doses of enu and mnu: While the chemicals differ dramatically in their mutagenicity in stem-cell spermatogonia, both elicit very high mutation rates in differentiating spermatogonia. *Mutat Res* 2007; **616**: 181
289. Russell LB, Hunsicker PR, Kerley MK et al: Bleomycin, unlike other male-mouse mutagens, is most effective in spermatogonia, inducing primarily deletions. *Mutat Res* 2000; **469**: 95
290. Yoshimoto Y, Neel JV, Schull WJ et al: Malignant tumors during the first 2 decades of life in the offspring of atomic bomb survivors. *Am J Hum Genet* 1990; **46**: 1041
291. Winther JF, Boice JD, Jr., Mulvihill JJ et al: Chromosomal abnormalities among offspring of childhood-cancer survivors in denmark: A population-based study. *Am J Hum Genet* 2004; **74**: 1282
292. Signorello LB, Mulvihill JJ, Green DM et al: Congenital anomalies in the children of cancer survivors: A report from the childhood cancer survivor study. *J Clin Oncol* 2012; **30**: 239
293. Al-Jebari Y, Glimelius I, Berglund Nord C et al: Cancer therapy and risk of congenital malformations in children fathered by men treated for testicular germ-cell cancer: A nationwide register study. *PLoS Med* 2019; **16**: e1002816
294. Robbins WA, Meistrich ML, Moore D et al: Chemotherapy induces transient sex chromosomal and autosomal aneuploidy in human sperm. *Nat Genet* 1997; **16**: 74
295. Monteil M, Rousseaux S, Chevret E et al: Increased aneuploid frequency in spermatozoa from a hodgkin's disease patient after chemotherapy and radiotherapy. *Cytogenet Cell Genet* 1997; **76**: 134
296. Martin RH, Ernst S, Rademaker A et al: Chromosomal abnormalities in sperm from testicular cancer patients before and after chemotherapy. *Hum Genet* 1997; **99**: 214
297. De Mas P, Daudin M, Vincent MC et al: Increased aneuploidy in spermatozoa from testicular tumour patients after chemotherapy with cisplatin, etoposide and bleomycin. *Hum Reprod* 2001; **16**: 1204
298. Martinez G, Walschaerts M, Le Mitouard M et al: Impact of hodgkin or non-hodgkin lymphoma and their treatment^{ts} on sperm aneuploidy: A prospective study by the french cecos network. *Fertil Steril* 2017; **107**: 341
299. Martin RH, Ernst S, Rademaker A et al: Analysis of sperm chromosome complements before, during, and after chemotherapy. *Cancer Genet Cytogenet* 1999; **108**: 133

300. Bogefors K, Giwercman YL, Eberhard J et al: Androgen receptor gene cag and ggn repeat lengths as predictors of recovery of spermatogenesis following testicular germ cell cancer treatment. *Asian J Androl* 2017; **19**: 538
301. Bujan L, Walschaerts M, Moinard N et al: Impact of chemotherapy and radiotherapy for testicular germ cell tumors on spermatogenesis and sperm DNA: A multicenter prospective study from the cecos network. *Fertil Steril* 2013; **100**: 673
302. O'Flaherty C, Hales BF, Chan P and Robaire B: Impact of chemotherapeutics and advanced testicular cancer or hodgkin lymphoma on sperm deoxyribonucleic acid integrity. *Fertil Steril* 2010; **94**: 1374
303. Di BC, Bertagna A, Composto ER et al: Effects of oncological treatments on semen quality in patients with testicular neoplasia or lymphoproliferative disorders. *Asian J Androl* 2013; **15**: 425
304. Kawai K and Nishiyama H: Preservation of fertility of adult male cancer patients treated with chemotherapy. *Int J Clin Oncol* 2019; **24**: 34
305. Oktay K, Harvey BE, Partridge AH et al: Fertility preservation in patients with cancer: Asco clinical practice guideline update. *J Clin Oncol* 2018; **36**: 1994
306. Fertility preservation in patients undergoing gonadotoxic therapy or gonadectomy: A committee opinion. *Fertil Steril* 2019; **112**: 1022
307. Hsiao W, Deveci S and Mulhall JP: Outcomes of the management of post-chemotherapy retroperitoneal lymph node dissection-associated anejaculation. *BJU Int* 2012; **110**: 1196
308. Berookhim BM and Mulhall JP: Outcomes of operative sperm retrieval strategies for fertility preservation among males scheduled to undergo cancer treatment. *Fertil Steril* 2014; **101**: 805
309. Ombelet W, Dhont N, Thijssen A et al: Semen quality and prediction of iui success in male subfertility: A systematic review. *Reprod Biomed Online* 2014; **28**: 300
310. Lemmens L, Kos S, Beijer C et al: Predictive value of sperm morphology and progressively motile sperm count for pregnancy outcomes in intrauterine insemination. *Fertil Steril* 2016; **105**: 1462
311. Nangia AK, Luke B, Smith JF et al: National study of factors influencing assisted reproductive technology outcomes with male factor infertility. *Fertil Steril* 2011; **96**: 609
312. Williams DHT, Karpman E, Sander JC et al: Pretreatment semen parameters in men with cancer. *J Urol* 2009; **181**: 736
313. Agarwal A, Shekarriz M, Sidhu RK and Thomas AJ, Jr.: Value of clinical diagnosis in predicting the quality of cryopreserved sperm from cancer patients. *J Urol* 1996; **155**: 934
314. Auger J, Sermondade N and Eustache F: Semen quality of 4480 young cancer and systemic disease patients: Baseline data and clinical considerations. *Basic Clin Androl* 2016; **26**: 3
315. Grover NS, Deal AM, Wood WA and Mersereau JE: Young men with cancer experience low referral rates for fertility counseling and sperm banking. *J Oncol Pract* 2016; **12**: 465
316. Klosky JL, Wang F, Russell KM et al: Prevalence and predictors of sperm banking in adolescents newly diagnosed with cancer: Examination of adolescent, parent, and provider factors influencing fertility preservation outcomes. *J Clin Oncol* 2017; **35**: 3830
317. Sonnenburg DW, Brames MJ, Case-Eads S and Einhorn LH: Utilization of sperm banking and barriers to its use in testicular cancer patients. *Support Care Cancer* 2015; **23**: 2763
318. Bizet P, Saias-Magnan J, Jouve E et al: Sperm cryopreservation before cancer treatment: A 15-year monocentric experience. *Reprod Biomed Online* 2012; **24**: 321
319. van Casteren NJ, van Santbrink EJ, van Inzen W et al: Use rate and assisted reproduction technologies outcome of cryopreserved semen from 629 cancer patients. *Fertil Steril* 2008; **90**: 2245
320. Ferrari S, Paffoni A, Filippi F et al: Sperm cryopreservation and reproductive outcome in male cancer patients: A systematic review. *Reprod Biomed Online* 2016; **33**: 29
321. O'Flaherty CM, Chan PT, Hales BF and Robaire B: Sperm chromatin structure components are differentially repaired in cancer survivors. *J Androl* 2012; **33**: 629

322. Weibring K, Nord C, StÅhl O et al: Sperm count in swedish clinical stage i testicular cancer patients following adjuvant treatment. *Ann Oncol* 2019; **30**: 604
323. Anserini P, Chiodi S, Spinelli S et al: Semen analysis following allogeneic bone marrow transplantation. Additional data for evidence-based counselling. *Bone Marrow Transplant* 2002; **30**: 447
324. Rives N, Walschaerts M, Setif V et al: Sperm aneuploidy after testicular cancer treatment: Data from a prospective multicenter study performed within the french centre d'a%tude et de conservation des oeufs et du sperme network. *Fertil Steril* 2017; **107**: 580
325. StÅhl O, Eberhard J, Jepson K et al: Sperm DNA integrity in testicular cancer patients. *Hum Reprod* 2006; **21**: 3199
326. Suzuki K, Yumura Y, Ogawa T et al: Regeneration of spermatogenesis after testicular cancer chemotherapy. *Urol Int* 2013; **91**: 445
327. Alwaal A, Breyer BN and Lue TF: Normal male sexual function: Emphasis on orgasm and ejaculation. *Fertil Steril* 2015; **104**: 1051
328. Jacobsen KD, Ous S, Waehre H et al: Ejaculation in testicular cancer patients after post-chemotherapy retroperitoneal lymph node dissection. *Br J Cancer* 1999; **80**: 249
329. Meseguer M, Garrido N, RemohA J et al: Testicular sperm extraction (tese) and icsi in patients with permanent azoospermia after chemotherapy. *Hum Reprod* 2003; **18**: 1281
330. Hsiao W, Stahl PJ, Osterberg EC et al: Successful treatment of postchemotherapy azoospermia with microsurgical testicular sperm extraction: The weill cornell experience. *J Clin Oncol* 2011; **29**: 1607
331. Dar S, Orvieto R, Levron J et al: Ivf outcome in azoospermic cancer survivors. *Eur J Obstet Gynecol Reprod Biol* 2018; **220**: 84
332. Shin T, Kobayashi T, Shimomura Y et al: Microdissection testicular sperm extraction in japanese patients with persistent azoospermia after chemotherapy. *Int J Clin Oncol* 2016; **21**: 1167
333. Shiraishi K and Matsuyama H: Microdissection testicular sperm extraction and salvage hormonal treatment in patients with postchemotherapy azoospermia. *Urology* 2014; **83**: 100
334. Zorn B, Virant-Klun I, Stanovnik M et al: Intracytoplasmic sperm injection by testicular sperm in patients with aspermia or azoospermia after cancer treatment. *Int J Androl* 2006; **29**: 521
335. Chan PTK, Palermo GD, Veeck LL et al: Testicular sperm extraction combined with intracytoplasmic sperm injection in the treatment of men with persistent azoospermia postchemotherapy. *Cancer* 2001; **92**: 1632
336. Tannour-Louet M, Han S, Corbett ST et al: Identification of de novo copy number variants associated with human disorders of sexual development. *PLoS One* 2010; **5**: e15392
337. Tannour-Louet M, Han S, Louet JF et al: Increased gene copy number of vamp7 disrupts human male urogenital development through altered estrogen action. *Nat Med* 2014; **20**: 715
338. Haller M, Au J, O'Neill M and Lamb DJ: 16p11.2 transcription factor maz is a dosage-sensitive regulator of genitourinary development. *Proc Natl Acad Sci U S A* 2018; **115**: E1849
339. Haller M, Mo Q, Imamoto A and Lamb DJ: Murine model indicates 22q11.2 signaling adaptor crkl is a dosage-sensitive regulator of genitourinary development. *Proc Natl Acad Sci U S A* 2017; **114**: 4981
340. Jorgez CJ, Rosenfeld JA, Wilken NR et al: Genitourinary defects associated with genomic deletions in 2p15 encompassing otx1. *PLoS One* 2014; **9**: e107028
341. Pryor JL, Kent-First M, Muallem A et al: Microdeletions in the y chromosome of infertile men. *N Engl J Med* 1997; **336**: 534
342. Vogt P, Chandley AC, Hargreave TB et al: Microdeletions in interval 6 of the y chromosome of males with idiopathic sterility point to disruption of azf, a human spermatogenesis gene. *Hum Genet* 1992; **89**: 491
343. Ma K, Sharkey A, Kirsch S et al: Towards the molecular localisation of the azf locus: Mapping of microdeletions in azoospermic men within 14 subintervals of interval 6 of the human y chromosome. *Hum Mol Genet* 1992; **1**: 29

344. Genecards®: The human gene database: GeneCards, vol. 2020
345. Coutton C, Escoffier J, Martinez G et al: Teratozoospermia: Spotlight on the main genetic actors in the human. Hum Reprod Update 2015; **21**: 455
346. Wang WL, Tu CF and Tan YQ: Insight on multiple morphological abnormalities of sperm flagella in male infertility: What is new? Asian J Androl 2020; **22**: 236
347. Matzuk MM and Lamb DJ: The biology of infertility: Research advances and clinical challenges. Nat Med 2008; **14**: 1197
348. Matzuk MM and Lamb DJ: Genetic dissection of mammalian fertility pathways. Nat Cell Biol 2002; **4 Suppl**: s41
349. Oud MS, Volozonoka L, Smits RM et al: A systematic review and standardized clinical validity assessment of male infertility genes. Hum Reprod 2019; **34**: 932
350. Kasturi SS, Tannir J and Brannigan RE: The metabolic syndrome and male infertility. J Androl 2008; **29**: 251
351. Hehemann MC, Raheem OA, Rajanahally S et al: Evaluation of the impact of marijuana use on semen quality: A prospective analysis. Ther Adv Urol 2021; **13**: 17562872211032484
352. Morrison CD and Brannigan RE: Metabolic syndrome and infertility in men. Best Pract Res Clin Obstet Gynaecol 2015; **29**: 507
353. Eisenberg ML, Kim S, Chen Z et al: The relationship between male bmi and waist circumference on semen quality: Data from the life study. Hum Reprod 2014; **29**: 193
354. Brinster RL: Germline stem cell transplantation and transgenesis. Science 2002; **296**: 2174
355. Komeya M, Sato T and Ogawa T: In vitro spermatogenesis: A century-long research journey, still half way around. Reprod Med Biol 2018; **17**: 407
356. Kubota H and Brinster RL: Spermatogonial stem cells. Biol Reprod 2018; **99**: 52

Attention: Restrictions on use of AUA, AUAER, and UCF content in third party applications, including artificial intelligence technologies, such as large language models and generative AI.

You are prohibited from using or uploading content you accessed through this website into external applications, bots, software, or websites, including those using artificial intelligence technologies and infrastructure, including deep learning, machine learning and large language models and generative AI.



American Urological Association

1000 Corporate Boulevard
Linthicum, MD 21090
Phone: 410-689-3700
Toll-Free: 1-800-828-7866
Fax: 410-689-3912
Email: aua@AUAnet.org

About AUA

[AUA Overview](#)
[Annual Report](#)
[Mission & Vision](#)
[Leadership](#)
[Governance](#)
[Careers at the AUA](#)

Quick Links

[AUA Sections](#)
[Contact Us](#)
[Donate](#)
[Press/Media](#)
[Privacy Policy](#)
[Terms and Conditions of Use](#)
[Industry Relations](#)
[Interest-Based Advertising](#)

©2026 American Urological Association | All Rights Reserved.